

Invertebrates as a study model of anaerobic infections

Nevretenčarski modeli za proučevanje anaerobnih infekcij

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Abstract: Experiments with invertebrates have recently gained increased attention as a practicable substitute to traditional mammalian models in the study of host-bacterial interactions. Using an invertebrate study model has a number of advantages over traditional mammalian model including simple growth condition, short life-time, can be easily maintained, infected without anesthesia and with a smaller extent of ethical limitations. From a microbiological viewpoint, importance of anaerobic bacteria as agents for various diseases remains an interesting field for research. The study of the interaction between invertebrate model host and anaerobic bacteria therefore provides insights into the mechanisms underlying pathogen virulence and host immunity and complements or even compensates the use of mammalian model in assay for infectious disease. This review offers to consider about the appropriate invertebrate model select for the study of particular aspects of anaerobic bacterial pathogenesis.

Keywords: invertebrate model, anaerobic bacteria, virulence factors, disease

Izvilleček: Poskusi na nevretenčarjih so lahko odlični nadomestni model za proučevanje interakcij med gostiteljem in bakterijo. Uporaba nevretenčarjev ima številne prednosti pred uporabo sesalskih živalskih modelov za raziskovalne poskuse. Pogoji za življenje so enostavni, življenjska doba je kratka, vzdrževanje je enostavno, izvajanje poskusov ne vključuje anestezije, kot tudi uporaba nevretenčarjev za raziskovalne namene je manj etično sporna. Anaerobne bakterije in bolezni, ki jih povzročajo, so vedno zanimivo področje raziskovanja. Novi pristopi pri proučevanju možnih negativnih učinkov anaerobnih bakterij in virulenčnih dejavnikov bi lahko postali tudi nevretenčarski modeli. Prispevek opisuje uporabnost nevretenčarskih modelov, že opisane povezave med patogenimi učinki virulenčnih dejavnikov anaerobnih bakterij pri nevretenčarskih modelih in o novih pristopih izbire nevretenčarjev kot model za proučevanje patogeneze.

Ključne besede: nevretenčarski modeli, anaerobne bakterije, virulenčni dejavniki, bolezen

Introduction

The molecular basis of the pathogenicity of infectious agents, and of the corresponding mechanisms of host defence can be studied using model systems (Couillault and Ewbank 2002). There is a continuing need for the development of a simple animal model for the study of host pathogen interactions (Finlay 1999).

A number of different invertebrate host model systems have been described in the past few years that allow multidisciplinary studies of host–bacterial interactions from the perspectives of both the pathogen and the host. Consequently, many researchers have turned to invertebrates as effortless, practicable, simple, and inexpensive hosts to model a variety of human infectious diseases. It is important to select the model host that is best suited for testing a specific hypothesis (Mylonakis et al. 2007) including ethical, procedural and financial characteristics.

A number of different model systems, including amoeba, nematodes, crustaceans and insects, have been introduced, and it was observed that different bacteria responded in different ways to presumptive alternate hosts, and specific model systems might be more or less advisable for a defined pathogen (Ott et al. 2012). Aerobic and obligate anaerobic bacteria are successfully isolated but to isolate anaerobic bacteria from invertebrate models is often impracticable (Bergan 1984), even though, anaerobic bacteria in invertebrates cannot be excluded. The subsequent isolation of strictly anaerobic bacteria resulting in anaerobic microniches within an oxic environment. This finding of anaerobic microniches presents microhabitat of various microbes, despite the fact of its obviously inappropriate environment (König 2006).

This review presents five model hosts, the amoeba *Acanthamoeba polyphaga*, the nematode *Caenorhabditis elegans* (*C. elegans*), the fruit fly *Drosophila melanogaster* (*D. melanogaster*), the greater wax moth *Galleria mellonella* (*G. mellonella*) and the isopod *Porcellio scaber* (*P. scaber*). Predominant empirical advantages of each model are well developed genetic, biochemical and biological functions, the precision evaluation of an assay, conserved innate immune response, handling experiments at 37°C and ease of inoculation of an explicit amount of pathogen (Borner 2016).

Anaerobic bacteria are significant clinical pathogens

Anaerobes and their pathogenicity factors can affect common hosts and hosts with compromised resistance of harmed tissue. Their complex metabolism, the capability to produce pathogenicity components like extracellular toxins, superoxide dismutase, catalase, the abscess inducing capsular polysaccharide, proteases, lipases, heparinase, nucleases hyaluronidase, haemolysin, lipolysin, and neuroaminidase, enzymes inactivating antibiotics, and resistance against phagocytosis, are responsible for local and systemic expansion of the endogenous bacterial infection during antimicrobial therapy (Bergan 1984, Brook 2011, Dorer and Isberg 2006). Avoidance and early healing treatment of circumstances that can lead to anaerobic infection can reduce their amount.

Anaerobic bacteria are found on the skin, on mucosal surfaces, in the mouth, pharynx and intestinal tract or genital tract as a part of the normal microbiota. Additionally, anaerobes can be isolated in all types of anaerobic infection including respiratory infection, subcutaneous and soft-tissue infections, endogenous infections in the central nervous system, oral cavity, head and neck, chest, abdomen, pelvis, skin, and soft tissue (Brook 2016). Infections results when anaerobes and other bacteria of the normal flora weaken and deceive immune system to avoid detection (Borner 2016), or permeate integumentary barriers. The infections are often polymicrobial, with other anaerobes, facultative anaerobes, and aerobes (Brook 2016). Several important diseases, botulism, tetanus, gas gangrene, food poisoning, and pseudomembranous colitis are caused by anaerobic *Clostridium* species from the environment or from normal flora (Brook 2016, Brooks et al. 2010). Their individual pathogenicity factors serum-independent chemotactic factors that attract polymorphonuclear cells, superoxide dismutase, catalase, capsular structures, proteases, lipases, heparinase, and nucleases, exotoxins of histotoxic clostridia and the ability of different anaerobes to produce enzymes inactivating antibiotics has become well established. Pathogenicity factors like hyaluronidase, haemolysin, lipolysin, and neuroaminidase have been isolated (Brooks et al. 2010). In spite of all that, the importance of

other virulence factors that may contribute to the pathogenicity of the anaerobic bacteria remains unclear (Brooks et al. 2010, Harding et al. 2013).

Important anaerobes that may cause human infection and/or are isolated in polymicrobial anaerobic infections are: i. Gram-negative bacilli *Bacteroides* spp., *Prevotella* spp., *Porphyromonas* spp., *Fusobacterium* spp., *Bilophila* spp. and *Sutterella* spp., ii. Gram-negative cocci mainly *Veillonella* spp. (Brook, 2011), iii. Gram-positive cocci *Peptostreptococcus* spp., *Anaerococcus* spp., *Finexgoldia* spp., *Parvimonas* spp., and *Peptoniphilus* spp. (Murphy and Frick, 2013), iv. Gram-positive spore forming *Clostridium* spp., and no spore-forming bacilli *Actinomyces* spp., *Propionibacterium* spp., *Eubacterium* spp., *Lactobacillus* spp., *Bifidobacterium* spp., *Eggerthella* spp., *Arachnia* spp. (Brook 2011). The isolation and identification of anaerobic bacteria associated with specific infection are important as well as characterization of their virulence factors (Brook 2016, Murphy and Frick 2013).

Invertebrate as a model host for studying virulence of anaerobic bacteria

Invertebrate models have gained increased attention as a viable alternative to traditional mammalian models of infection (Mowlds et al. 2008, Renwick et al. 2006) and are increasingly being used to study a number of important human pathogens. Using of invertebrate models have number of advantages over traditional mammalian models, as invertebrates are not subject to the ethical limitations of mammalian models (Harding et al. 2013). No invertebrate model hosts reproduces all aspects of mammalian infection and any particular invertebrate is likely to have specific advantages. The selection of a model system for studying virulence of anaerobic bacteria is largely dependent on the specific pathogen virulence related factors, the specific host innate immune responses of interest, and the scientific question asked. If the goal is to study innate immune responses, the choice most likely will require the selection of a multicellular model genetic organism such as *D. melanogaster* or *C. elegans*. If the goal is to study phagocytosis and/or the outcome of ingestion, the choices

include unicellular organisms such as amoebae and slime mold or invertebrates such as insects with phagocytic cells (Mylonakis et al. 2007). If the goal is to study gut microbe homeostasis and gut infection by the human pathogen anaerobic bacteria, a model with increasing evidence for a reciprocal relationship between beneficial and pathogenic bacteria in the gut and the intestinal immune system with suitable environment for developing of resident and anaerobic microbiota must be found and practiced (Glavis-Bloom et al. 2012).

Manipulating with alimentary, physiological and behavioral characteristics of different invertebrate models might play an important role with an optimal adaptation of anaerobic bacteria to invertebrate's environment through a completely different mechanism of interactions ranging from pathogenesis to obligate mutualism.

Acanthamoeba polyphaga as a study model for anaerobic bacteria

Protozoa are frequently used in laboratories as experimental organisms for studies of cell locomotion (*Amoeba proteus*), nonmuscle contractile systems (*Acanthamoeba*), and the effects of removing and transplanting nuclei (Brusca and Brusca 2004).

Amoebae species are well established model systems for a number of pathogenic bacteria. Amoebae have been used as model organism to study the pathogenicity of bacterial strain, such as *Pseudomonas aeruginosa* (Pukatzki et al. 2002), as a biological tool for isolation of several amoeba-resisting intracellular microorganisms (Adekambi et al. 2004, Greub et al., 2004, La Scola et al. 2004) but it has not been an appropriate study model for anaerobic bacteria yet. *Clostridium frigidicarnis* was demonstrated to be lytic for amoebae (Pagnier et al. 2008).

Caenorhabditis elegans as a study model for anaerobic bacteria

The soil-living small size nematode *C. elegans* with rapid life cycle and transparent body, fully sequenced genome, and physiological and anatomical simplicity is a model host with excellent potential for studying cell biology and pathogenicity

(Brusca and Brusca 2004, Glavis-Bloom et al. 2012). Both, aerobic and anaerobic metabolic pathways are found and worm is able to switch from one pathway to the other according to environmental oxygen concentrations. Facultative anaerobiosis is evidently meaningful in parasitic nematodes and those that live in additional anoxic environments (Brusca and Brusca 2004).

Bacteria that infect *C. elegans* are both Gram-negative and Gram-positive bacteria. *Salmonella typhimurium*, *Aeromonas hydrophila*, *Pseudomonas aeruginosa*, *Burkholderia pseudomallei*, *Burkholderia cepacia*, *Burkholderia cenocepacia*, *Yersinia pestis*, *Yersinia pseudotuberculosis* are Gram-negative bacteria that infect *C. elegans* and *Staphylococcus aureus*, *Streptococcus pyogenes*, *Streptococcus pneumoniae*, *Microbacterium nematophilum*, *Enterococcus faecalis*, *Bacillus thuringiensis* are Gram-positive bacteria (Borner 2016).

Practical advantages, the particular bioinformatics approach and biological processes have increased the use of *C. elegans* in toxicological studies (Boyd et al. 2012). *C. elegans* is killed by many pathogens and many virulence factors produced by *pathogens* that contribute to the pathogenicity in humans have been shown to be important for disease in *C. elegans*, including persistent infection of the intestine, colonisation with biofilm formation on the worm cuticle, and killing by botulinum toxin, hydrogen cyanide or hydrogen peroxide (Kaletta and Hengartner 2006).

The genetically tractable nematode *C. elegans* has been extensively used to study bacterial virulence and offers many advantages (Mahajan-Miklos et al. 1999) as a convenient host for studies of pathogen infections (Balla and Troemel 2013). This small hermaphroditic animal has been the object of intense study for more than 20 years (Brillard 2001). From a microbiological standpoint, *C. elegans* is an attractive model for a broad range of host processes from the molecular level to the whole organism level. Genetic tractability and convenience, small size and simplicity, ease of culture, transparency and short lifespan all contribute to making *C. elegans* a useful laboratory organism for conducting large-scale studies of host-microbe interactions (Clark 2012). A large number of human diseases have been investigated using *C. elegans* (Kaletta and Hengartner 2006).

Several Gram-negative human pathogens have been shown to kill *C. elegans* when presented to the nematodes as a source of food (Aballay et al., 2000; Darby et al. 1999, Mahajan-Miklos et al. 1999, Tan et al. 1999). Gram-positive human pathogens also kill *C. elegans*. Literature is demonstrating that a range of aerobic and opportunistic bacterial pathogens is involved in virulence in *C. elegans* as well as an anaerobically grown *Enterococcus faecium*. *E. faecium* kills the nematode via the production of hydrogen peroxide, which also poses an oxidative stress to nematodes (Bolm et al. 2004, Borner 2016, Jansen et al. 2002, Moy et al. 2004). Notable research was made in 2014, where worms treated with botulinum toxin A of *Clostridium botulinum* showed slight paralysis and the toxin treatment resulted in the increase of yolk protein concentration in embryos (Kim et al. 2014). *C. elegans* has become an assisting model to probe vital biological and physiological processes and molecular mechanisms involved in many human diseases. It has served as a model for Parkinson's, Alzheimer's and Huntington's disease, diabetes, cancer, immune disorders, and the development and testing of therapeutics agents (Bier and McGinnis 2008, Wilson-Sanders 2011).

C. elegans may subsequently be the model of anaerobic microbial processes and toxicity.

Introducing bacteria to worms is quite simple, but laboratory standing conditions differ with the conditions in its natural soil habitat. The nematode lacks a variety of mammalian anatomical structures and the reproductive fitness of *C. elegans* strongly depends on the effects of air composition, habitat structure, and bacterial food availability. Owing to these not all diseases and immune responses can be testified (Glavis-Bloom et al. 2012) or have to be carefully interpreted by concern to their natural consequences (Freyth et al. 2010).

Galleria mellonella as a study model for anaerobic bacteria

Unique advantages of *G. mellonella* as a model host for studying pathogen virulence mechanisms and the efficiency of potential antimicrobial compounds are its ability to survive at 37°C when studying pathogenic temperature-sensitive virulence and production of microbial toxins. *G. mellonella* can be stored at room temperature,

likewise is straightforwardly and practically obtained in sizes large enough to be inoculated by pathogens (Glavis-Bloom et al. 2012). A reliable and inexpensive experimental infection model is convenient to differentiate between virulent and non-virulent isolates, for the identification of presumed virulence genes through comparative genomics studies and the identification of novel molecular targets for antimicrobial therapy and vaccine development. *G. mellonella* was recently established as a suitable host model to study the pathogenesis of bacterial and yeast species causing diseases in humans, such as *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Bacillus cereus*, *Cryptococcus neoformans*, and *Candida albicans* (Altincicek et al. 2012). Conversely, *G. mellonella* cannot replace well-established and more physiological *in vivo* experimental vertebrate models in the assessment of pathogenic mechanisms associated human diseases (Giannouli et al. 2014, Glavis-Bloom et al. 2012).

Drosophila melanogaster as a study model for anaerobic bacteria

The fruit fly *D. melanogaster* has many of the similar advantages as *C. elegans*. Small size, short generation time, a fully sequenced genome, and pre-existing libraries of genetic mutants, genes and pathways similar to those found in mammals completed *D. melanogaster* as an excellent model host (Frenzel et al. 2015). In particular, *D. melanogaster* is useful model for the study of mammalian intestinal bacterial infections (O'Callaghan and Vergunst 2010) and their effects on undifferentiated and matured enteric epithelial cells in the initial stages of intestinal cancer, for the study of intestinal infection with *Pseudomonas aeruginosa*, a human opportunistic bacterial pathogen (Glavis-Bloom et al. 2012) and as a model for the innate immune response to pathogens (Borner 2016) such as antimicrobial peptide production, phagocytosis and melanization reactions (Charroux and Royet 2012). Pathogens that infect *D. melanogaster* are *Salmonella* Typhimurium, *Pseudomonas aeruginosa*, *Mycobacterium marinum*, *Mycobacterium fortuitum*, *Staphylococcus aureus*, *Serratia marcescens*, and *Vibrio cholerae* (Borner 2016, O'Callaghan and Vergunst 2010).

The fruit fly *D. melanogaster* is an excellent model organism to research intestinal homeostasis, the gut microbiota, pathways that regulate intestinal stem cell signaling, innate immune reactions and disease. About 20 diverse species, including main dominant species of the genus *Acetobacter* and *Lactobacillus* and with few anaerobes were found in the intestine of *D. melanogaster*. No noteworthy anaerobic bacteria had been found in the *D. melanogaster* gastrointestinal tract (Charroux and Royet 2012) before the anaerobe *Clostridium perfringens* was isolated. The promoting effect on the growth and development of *D. melanogaster* were detected (Wei et al. 2016). It is a primary organism used in developing biology (Wilson-Sanders 2011) considering that *D. melanogaster* indirect-flight-muscle actin was ADP-ribosylated by *Clostridium botulinum* C2 toxin and *Clostridium perfringens* iota toxin (Just et al. 1993).

C. perfringens is the most frequently isolated histotoxic clostridia and produces several necrotizing extracellular toxins correlated with tissue necrosis, hemolytic anemia and renal failure and *C. botulinum* infections can result in intestinal toxemia, food poisoning and wound infections containing a highly potent neurotoxin (Brook 2016). Appropriate management of *D. melanogaster* for identifying and understanding anaerobes that are presented in human diseases could contribute to biological discoveries.

Drosophila served as a model for host-parasite relationships, is established model for the study of neoplastic diseases (Tipping and Perrimon 2014, Wang et al. 2014, Wilson-Sanders 2011), has been used to study cellular defenses against fungal pathogens (Arvanitis et al. 2013, Fuchs and Mylonakis 2006), it has been proven as excellent system for studying the normal function of human genes and pathways linked to neurodegenerative diseases (Allan et al. 2014, Liang et al. 2013, Mhatre et al. 2013), and is a simple model organism for studying diseases caused by viruses (Bier and Guichard 2012, Panayidou et al. 2014).

Porcellio scaber as a study model for anaerobic bacteria

The terrestrial crustacean, soil dwelling isopod *Porcellio scaber* is likely to be dependant on microorganisms associated with the gut (Kostanjšek

et al. 2004, Wang et al. 2004, Zimmer 2002). The digestive tract is complete with a well-developed, cuticle-lined, stomodeal foregut and proctodeal hindgut, connected by an entodermally derived midgut. A characteristic feature is a permeable peritrophic membrane to protect the delicate midgut epithelium from abrasion (Bier and McGinnis 2008).

Terrestrial isopods have an inherent and multiplicity gut microflora (Drobne 1995). Therefore, slight changes in the animal fitness might have effects on the microbial community in the intestinal tract (Guarner and Malagelada 2003; Loker et al. 2004). Due to their significant environmental aspect and their complete digestive tract a considerable amount of experimentation was focused on *P. scaber*. Anaerobic bacteria from *P. scaber* hindgut were identified. Further, obligate anaerobic bacteria of genus *Bacteroides* and *Enterococcus* species were isolated. Additionally, bacteria from the genus *Desulfotomaculum* were isolated from gut wall and cultivated under anaerobic conditions (Kostanjšek et al. 2004). Bacteria from the genus *Desulfotomaculum* were isolated from gut wall and cultivated under anaerobic conditions (Kostanjšek et al. 2004). Nothing can be concluded about the changed structure or function of the entire gut bacterial community. Gut microflora toxicity studies are a promising way to get applicable facts on terrestrial environments.

Conclusion

Importance of invertebrates as reservoirs of multihost pathogens often plays a crucial role of various infections. The invertebrates appeared for a long time to be an unsuitable environment for growth of anaerobic bacteria. The finding of anaerobic bacteria reveals that a unique microbial environment remains an interesting field for further microbiological research (König 2006) focusing on genotoxicity and other virulence factors, inflammation, host defences modulation, and bacterial derived metabolism. A better understanding of the interactions between the invertebrate host and anaerobe pathogenicity depends on further functional studies and findings in almost every area of biology and medicine (Gagniere et al. 2016).

A very few anaerobic bacteria have been isolated in only two invertebrate models, *D. melanogaster* and *P. scaber*. Although anaerobic bacteria are unimpressive found in gastrointestinal tract of invertebrates, inflammatory diseases of the intestine arise from imbalanced interactions between the host gut epithelia and resident or ingested anaerobic microbes. Developing *in vivo* disease models with well characterized development and simple immunity (Chamilos et al. 2007) can help to explicate the basic mechanism underlying disease (Mhatre et al. 2013) start with a similarly important pathogenic role and life-threatening infections in invertebrates. The intestinal niche is also challenged continuously by numerous environmentally derived bacteria because of its exposed anatomy that is accessible to the external environment (Lee and Lee 2014).

Invertebrate model hosts represent valuable tools for the study of host-pathogen interactions because they facilitate the identification of bacterial virulence factors and allow the discovery of novel components involved in host innate immune responses (Miyata et al. 2003, König 2006). As well as facilitating the identification and study of virulence mechanisms (Mahajan-Miklos et al. 2000), simple model system may also permit direct genetic approaches for the study of host defenses (Ewbank 2002). The finding that diverse bacteria are pathogenic to invertebrate models opens the prospect of using this experimentally simple model to identify genes that are necessary not only for pathogenesis in study model but also for virulence or symbiosis in other hosts (Aballay et al. 2000, König 2006).

To study the pathogenesis in mammalian models is complicated by difficulties of handling, long reproductive cycles, small brood sizes, physiological and anatomical complexity, regulatory requirements, high cost, and ethical considerations. Workers in the field of pathogenesis have the opportunity to select from several invertebrate animal model systems in their studies. An understanding of the unique strengths and limitations associated with each model host is necessary, as particular virulence characteristics are not equally important in all systems and genetic tractability is not available in all model hosts (Mylonakis et al. 2007) and especially when a presence of the anaerobic pathogens is looking for. A better model

systems may be identified and fully characterized in the future, like feeding-based infections, to use pathogens that invade the luminal side of epithelial cells (Balla and Troemel 2013) or to study systemic infections by microbial injection into the hemolymph (Panayidou et al. 2014). By using and trying different experimental techniques and protocol details, observed the progression of infection in real-time by light microscopy, by fluorescence microscopy and by electron microscopy (Shu et al. 2011) and with well-established genetic, molecular and biochemical analyses of invertebrate animal models, a research model to facilitate maintenance of virulence by anaerobic bacteria could be created.

Genetic screening, the RNAi technique in the genetically tractable invertebrate model organisms have been proved to be a powerful and valuable tool for understanding of fundamental principles of bacterial resistance to infection and may be useful in screening for potential neurotoxicity (Abnave et al. 2015, Altincicek et al. 2007). Inflammatory diseases of the intestine, gastrointestinal cancer and gut-associated pathologies arise from imbalanced interactions between the host gut epithelia and resident or ingested microbes, interactions that are still poorly understood at the molecular level. *D. melanogaster* has been a very powerful model to study development and diseases (Charroux and Royet 2012).

The models reviewed are relatively inexpensive, easy to work with, have short lifespans, and often have very well characterized and stereotypical development and behaviour. Invertebrate models could serve as references for scientists concerned in alternatives to vertebrate animals (Lehner and Lee 2008) and could be a challenge for studying the pathogenesis of infections caused by anaerobic bacteria.

Povzetek

Preučevanje interakcij med nevretenčarji in anaerobnimi patogenimi bakterijami je na vseh področjih biologije in medicine pomembno, saj lahko vsaka odkrita medsebojna odvisnost pomembno vpliva na nastanek in razvoj bolezni (Gagniere in sod. 2016).

Število mikroorganizmov se pri nevretenčarjih nenehno spreminja, med drugim tudi zaradi okolja povezanega z gostiteljem, preproste anatomije in zato anaerobnih bakterij skoraj ni mogoče kultivirati (Lee in Lee, 2014). Teh mikrobov je malo. Prav izpostavljenost anaerobnim bakterijam pa poveča tveganje za kolonizacijo s sevi, ki so specifični za številne črevesne in izvenčrevesne okužbe. Zapis o izoliranih anaerobnih bakterijah so pri dveh nevretenčarskih modelih, *D. melanogaster* (Charroux in Royet 2012) in *P. scaber* (Kostanjšek 2004, König 2006).

Nevretenčarji so bili dolgo časa neprimerni za rast in razmnoževanje anaerobnih bakterij, saj organizem ne ustvarja pogojev zanje. Znanstveniki pa so z izolacijo anaerobnih bakterij pri nevretenčarjih potrdili, da se lahko zaradi prilagodljivosti v okolju ustvarijo tudi rastni pogoji za pritrđitev in razmnoževanje anaerobnih bakterij. Prav ta ugotovitev je vzbudila raziskovalce za nadaljevanje mikrobioloških raziskav v tem edinstvenem mikrobnem okolju nevretenčarjev (König 2006).

Dejavniki, ki vplivajo na mnoga bolezenska stanja izvirajo iz sestave črevesnih bakterij, njihovega vpliva na delovanje imunskega sistema in njegovega ravnovesja ter izražanje genov. Znanstveni pristopi pri raziskavah sestave anaerobnih bakterij pri nevretenčarjih so lahko v dosednji praksi delno uporabljeni za obvladovanje različnih bolezenskih sprememb (Mhatre in sod. 2013, Chamilos in sod. 2007). Prepoznavanje simptomov bolezni, poznavanje vrste in vloge anaerobov, dovzetnosti za posamezne bolezni so odraz na ravni gostitelja in mikroba. Njihov vpliv na fiziologijo črevesja, regulacijo metabolizma, razvoj in aktivacijo imunskega sistema so raziskovali v primerjalnih študijah na nevretenčarskih modelih (Aballay in sod. 2000, Miyata in sod. 2003, König 2006). Veliko je še neodkritega na tem področju. A z razvojem sodobnih molekularnih metod in eksperimentalnih tehnik (Shu in sod. 2011, Abnave in sod. 2015, Altincicek in sod. 2007) je mogoče prepoznavanje in opredeljevanje posamezne vrste bakterije in za preučevanje vloge genov so izjemno priročni tudi nevretenčarski modeli (Mahajan-Miklos in sod. 2000, Ewbank 2002, König 2006). Prav zaradi tega predstavljajo dragoceni model za preučevanje virulenčnih dejavnikov anaerobnih bakterij, saj omogočajo

tudi identifikacijo komponent, ki so vključene v imunski odziv (Aballay in sod. 2000).

Nevretenčarski modeli so enostavni za delo, njihova življenjska doba je kratka, izvajanje poskusov je ponovljivo in uporaba je manj etično sporna. Vsak nevretenčarski model ima poznane prednosti in slabosti ter poznane vse pomembne dejavnike, ki so lahko povezani z nastankom in razvojem bolezni (Mylonakis in sod. 2007, Balla in Troemel 2013, Panayidou in sod. 2014). Zato so lahko kot alternativa živalskim poskusom (Lehner in Lee 2008) in nov izziv v raziskovanju patogen-eze okužb, ki jih povzročajo anaerobne bakterije.

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Conflicts of interest

The authors had no conflicts of interest to declare in relation to this article.

References

- Aballay, A., Yorgey, P., Ausubel, F.M., 2000. *Salmonella typhimurium* proliferates and establishes a persistent infection in the intestine of *Caenorhabditis elegans*. *Curr. Biol.* 10(23), 1539-42.
- Abnave, P., Conti, F., Torre, C., Ghigo, E., 2015. What RNAi screens in model organisms revealed about microbicidal response in mammals? *Front. Cell Infect. Microbiol.* 4, 184.
- Adekambi, T., Reynaud-Gaubert, M., Greub, G., Gevaudan, M.J., La Scola, B., Raoult, D., Drancourt, M., 2004. Amoebal coculture of "*Mycobacterium massiliense*" sp. nov. from the sputum of a patient with hemoptoic pneumonia. *J. Clin. Microbiol.* 42(12), 5493-501.
- Allan, K., Perez, K.A., Barnham, K.J., Camakaris, J., Burke, R., 2014. A commonly used *Drosophila* model of Alzheimer's disease generates an aberrant species of amyloid-beta with an additional N-terminal glutamine residue. *FEBS Lett.* 588(20), 3739-43.
- Altincicek, B., Linder, M., Linder, D., Preissner, K.T., Vilcinskis, A., 2007. Microbial metalloproteinases mediate sensing of invading pathogens and activate innate immune responses in the lepidopteran model host *Galleria mellonella*. *Infect. Immun.* 75(1), 175-83.
- Arvanitis, M., Glavis-Bloom, J., Mylonakis, E., 2013. Invertebrate models of fungal infection. *Biochim. Biophys. Acta* 1832(9), 1378-83.
- Balla, K.M., Troemel, E.R., 2013. *Caenorhabditis elegans* as a model for intracellular pathogen infection. *Cell. Microbiol.* 15(8), 1313-22.
- Bergan, T., 1984. Pathogenicity of anaerobic bacteria. *Scand. J. Gastroenterol. Suppl* 91, 1-11.
- Bier, E., McGinnis, W., 2008. Model Organisms in the study of development and disease. In: Epstein, C.J., R.P. Erickson, A. Wynshaw-Boris (eds.): *Molecular Basis of Inborn Errors of Development*. Oxford University Press, New York, vol. 3, pp. 25-48.
- Bier, E., Guichard, A., 2012. Deconstructing host-pathogen interactions in *Drosophila*. *Dis. Model. Mech.* 5(1), 48-61.
- Bolm, M., Chhatwal, G.S., Jansen, W.T., 2004. Bacterial resistance of daf-2 mutants. *Science* 303(5666), 1976.
- Bolm, M., Jansen, W.T., Schnabel, R., Chhatwal, G.S., 2004. Hydrogen peroxide-mediated killing of *Caenorhabditis elegans*: a common feature of different streptococcal species. *Infect. Immun.* 72(2), 1192-4.
- Borner, R.A., 2016. Isolation and Cultivation of Anaerobes. *Adv. Biochem. Eng. Biotechnol.* 156, 35-53.
- Boyd, W.A., Smith, M.V., Freedman, J.H., 2012. *Caenorhabditis elegans* as a model in developmental toxicology. *Methods Mol. Biol.* 889, 15-24.
- Brillard, J., Ribeiro, C., Boemare, N., Brehelin, M., Givaudan, A., 2001. Two distinct hemolytic activities in *Xenorhabdus nematophila* are active against immunocompetent insect cells. *Appl. Environ. Microbiol.* 67(6), 2515-25.

- Brook, I., 2011. Antimicrobial treatment of anaerobic infections. *Expert. Opin. Pharmacother.* 12(11), 1691-707.
- Brook, I., 2016. Spectrum and treatment of anaerobic infections. *J. Infect. Chemother.* 22(1), 1-13.
- Brooks, G.F., Carroll, C.K., Butel, J.S., Morse, S.A., Mietzner, T.A., 2010. Jawetz, Melnick, & Adelberg's Medical Microbiology, ed. 25. McGraw-Hill, p. 814.
- Brusca, R.C., Brusca, G.J., 2004. Invertebrates. *Systematic Biology* 53(4), 662-664.
- Chamilos, G., Lionakis, M.S., Lewis, R.E., Kontoyiannis, D.P., 2007. Role of mini-host models in the study of medically important fungi. *Lancet Infect. Dis.* 7(1), 42-55.
- Charroux, B., Royet, J., 2012. Gut-microbiota interactions in non-mammals: what can we learn from *Drosophila*? *Semin. Immunol.* 24(1), 17-24.
- Clark, T.A., 2012. Responding to pertussis. *J. Pediatr.* 161(6), 980-2.
- Couillault, C., Ewbank, J.J., 2002. Diverse bacteria are pathogens of *Caenorhabditis elegans*. *Infect. Immun.* 70(8), 4705-7.
- Darby, C., Cosma, C.L., Thomas, J.H., 1999. Manoil Lethal paralysis of *Caenorhabditis elegans* by *Pseudomonas aeruginosa*. *Proc. Natl. Acad. Sci. USA* 96(26), 15202-7.
- Dorer, M.S., Isberg, R.R., 2006. Non-vertebrate hosts in the analysis of host-pathogen interactions. *Microbes Infect.* 8(6), 1637-46.
- Drobne, D., 1995. Bacteria adherent to the hindgut of terrestrial isopods. *Acta Microbiol. Immunol. Hung.* 42(1), 45-52.
- Ewbank, J.J., 2002. Tackling both sides of the host-pathogen equation with *Caenorhabditis elegans*. *Microbes Infect.* 4(2), 247-56.
- Finlay, B.B., 1999. Bacterial disease in diverse hosts. *Cell* 96(3), 315-8.
- Frenzel, E., Kranzler, M., Stark, T.D., Hofmann, T., Ehling-Schulz, M., 2015. The endospore-forming pathogen *Bacillus cereus* exploits a small colony variant-based diversification strategy in response to aminoglycoside exposure. *mBio* 6(6), e01172-15.
- Freyth, K., Janowitz, T., Nunes, F., Voss, M., Heinick, A., Bertaux, J., Scheu, S., Paul, R.J., 2010. Reproductive fitness and dietary choice behavior of the genetic model organism *Caenorhabditis elegans* under semi-natural conditions. *Mol. Cells* 30(4), 347-53.
- Fuchs, B.B., Mylonakis, E., 2006. Using non-mammalian hosts to study fungal virulence and host defense. *Curr. Opin. Microbiol.* 9(4), 346-51.
- Gagniere, J., Raisch, J., Veziat, J., Barnich, N., Bonnet, R., Buc, E., Bringer, M.A., Pezet, D., Bonnet, M., 2016. Gut microbiota imbalance and colorectal cancer. *World J. Gastroenterol.* 22(2), 501-18.
- Giannouli, M., Palatucci, A.T., Rubino, V., Ruggiero, G., Romano, M., Triassi, M., Ricci, V., Zarrilli, R., 2014. Use of larvae of the wax moth *Galleria mellonella* as an in vivo model to study the virulence of *Helicobacter pylori*. *BMC Microbiol.* 14, 228.
- Glavis-Bloom, J., Muhammed, M., Mylonakis, E., 2012. Of model hosts and man: using *Caenorhabditis elegans*, *Drosophila melanogaster* and *Galleria mellonella* as model hosts for infectious disease research. *Adv. Exp. Med. Biol.* 710, 11-7.
- Greub, G., La Scola, B., Raoult, D., 2004. Amoebae-resisting bacteria isolated from human nasal swabs by amoebal coculture. *Emerg. Infect. Dis.* 10(3), 470-7.
- Guarner, F., Malagelada, J.R., 2003. Gut flora in health and disease. *Lancet* 361(9356), 512-9.
- Harding, C.R., Schroeder, G.N., Collins, J.W., Frankel, G., 2013. Use of *Galleria mellonella* as a model organism to study *Legionella pneumophila* infection. *J. Vis. Exp.* 81, e50964.
- Hofstad, T., 1992. Virulence factors in anaerobic bacteria. *Eur. J. Clin. Microbiol. Infect. Dis.* 11(11), 1044-8.
- Jansen, W.T., Bolm, M., Balling, R., Chhatwal, G.S., Schnabel, R., 2002. Hydrogen peroxide-mediated killing of *Caenorhabditis elegans* by *Streptococcus pyogenes*. *Infect. Immun.* 70(9), 5202-7.
- Just, I., Hennessey, E.S., Drummond, D.R., Aktories, K., Sparrow, J.C., 1993. ADP-ribosylation of *Drosophila* indirect-flight-muscle actin and arthrin by *Clostridium botulinum* C2 toxin and *Clostridium perfringens* iota toxin. *Biochem. Journal* 291(2), 409-12.

- Kaletta, T., Hengartner, M.O., 2006. Finding function in novel targets: *C. elegans* as a model organism. *Nat. Rev. Drug Discov.* 5(5), 387-98.
- Kim, D.W., Lee, S.K., Ahnn, J., 2014. Phenotypic effect of botulinum toxin A on *Caenorhabditis elegans*. *Animal Cells Syst.* 18(3), 172-7.
- König, H., 2006. *Intestinal microorganisms of termites and other invertebrates*. Springer-Verlag Berlin Heidelberg, Germany.
- Kostanjšek, R., Lapanje, A., Rupnik, M., Štrus, J., Drobne, D., Avguštin, G., 2004. Anaerobic bacteria in the gut of terrestrial isopod Crustacean *Porcellio scaber*. *Folia microbiol.* 49(2), 179-82.
- La Scola, B., Birtles, R.J., Greub, G., Harrison, T.J., Ratcliff, R.M., Raoult, D., 2004. *Legionella drancourtii* sp. nov., a strictly intracellular amoebal pathogen. *Int. J. Syst. Evol. Microbiol.* 54(3), 699-703.
- Lee, K.A., Lee, W.J., 2014. *Drosophila* as a model for intestinal dysbiosis and chronic inflammatory diseases. *Dev. Comp. Immunol.* 42(1), 102-10.
- Lehner, B., Lee, I., 2008. Network-guided genetic screening: building, testing and using gene networks to predict gene function. *Brief Funct. Genomics Proteomics* 7(3), 217-27.
- Liang, J., Luo, J., Jin, J., 2013. Study of Parkinson's disease based on *Drosophila* model. *J. Zhejiang University, Medical Sciences* 42(6), 685-92.
- Loker, E.S., Adema, C.M., Zhang, S.M., Kepler, T.B., 2004. Invertebrate immune systems-not homogeneous, not simple, not well understood. *Immunol. Rev.* 198, 10-24.
- Mahajan-Miklos, S., Rahme, L.G., Ausubel, F.M., 2000. Elucidating the molecular mechanisms of bacterial virulence using non-mammalian hosts. *Mol. Microbiol.* 37(5), 981-8.
- Mahajan-Miklos, S., Tan, M.W., Rahme, L.G., Ausubel, F.M., 1999. Molecular mechanisms of bacterial virulence elucidated using a *Pseudomonas aeruginosa*-*Caenorhabditis elegans* pathogenesis model. *Cell* 96(1), 47-56.
- Mhatre, S.D., Paddock, B.E., Saunders, A.J., Marena, D.R., 2013. Invertebrate models of Alzheimer's disease. *J. Alzheimers Dis.* 33(1), 3-16.
- Miyata, S., Casey, M., Frank, D.W., Ausubel, F.M., Drenkard, E., 2003. Use of the *Galleria mellonella* caterpillar as a model host to study the role of the type III secretion system in *Pseudomonas aeruginosa* pathogenesis. *Infect. Immun.* 71(5), 2404-13.
- Mowlds, P., Barron, A., Kavanagh, K., 2008. Physical stress primes the immune response of *Galleria mellonella* larvae to infection by *Candida albicans*. *Microbes Infect.* 10(6), 628-34.
- Moy, T.I., Mylonakis, E., Calderwood, S.B., Ausubel, F.M., 2004. Cytotoxicity of hydrogen peroxide produced by *Enterococcus faecium*. *Infect. Immun.* 72(8), 4512-20.
- Murphy, E.C., Frick, I.M., 2013. Gram-positive anaerobic cocci-commensals and opportunistic pathogens. *FEMS Microbiol. Rev.* 37(4), 520-53.
- Mylonakis, E., Casadevall, A., Ausubel, F.M., 2007. Exploiting amoeboid and non-vertebrate animal model systems to study the virulence of human pathogenic fungi. *PLoS Pathog.* 3(7), e101.
- O'Callaghan, D., Vergunst, A., 2010. Non-mammalian animal models to study infectious disease: worms or fly fishing? *Curr. Opin. Microbiol.* 13(1), 79-85.
- Ott, L., McKenzie, A., Baltazar, M.T., Britting, S., Bischof, A., Burkovski, A., 2012. Evaluation of invertebrate infection models for pathogenic corynebacteria. *FEMS Immunol. Med. Microbiol.* 65(3), 413-21.
- Pagnier, I., Raoult, D., La Scola, B., 2008. Isolation and identification of amoeba-resisting bacteria from water in human environment by using an *Acanthamoeba polyphaga* co-culture procedure. *Environ. Microbiol.* 10(5), 1135-44.
- Panayidou, S., Ioannidou, E., Apidianakis, Y., 2014. Human pathogenic bacteria, fungi, and viruses in *Drosophila*: disease modeling, lessons, and shortcomings. *Virulence* 5(2), 253-69.
- Pukatzki, S., Kessin, R.H., Mekalanos, J.J., 2002. The human pathogen *Pseudomonas aeruginosa* utilizes conserved virulence pathways to infect the social amoeba *Dictyostelium discoideum*. *Proc. Natl. Acad. Sci. USA* 99(5), 3159-64.

- Renwick, J., Daly, P., Reeves, E.P., Kavanagh, K., 2006. Susceptibility of larvae of *Galleria mellonella* to infection by *Aspergillus fumigatus* is dependent upon stage of conidial germination. *Mycopathologia* 161(6), 377-84.
- Shu, X., Lev-Ram, V., Deerinck, T.J., Qi, Y., Ramko, E.B., Davidson, M.W., Jin, Y., Ellisman, M.H., Tsien, R.Y., 2011. A genetically encoded tag for correlated light and electron microscopy of intact cells, tissues, and organisms. *PLoS Biol.* 9(4), e1001041.
- Tan, M.W., Mahajan-Miklos, S., Ausubel, F.M., 1999. Killing of *Caenorhabditis elegans* by *Pseudomonas aeruginosa* used to model mammalian bacterial pathogenesis. *Proc. Natl. Acad. Scil USA* 96(2), 715-20.
- Tipping, M., Perrimon, N., 2014. *Drosophila* as a model for context-dependent tumorigenesis. *Jl Cell Physioll* 229(1), 27-33.
- Wang, L., Kounatidis, I., Ligoxygakis, P., 2014. *Drosophila* as a model to study the role of blood cells in inflammation, innate immunity and cancer. *Front. Cell Infect. Microbiol.* 3, 113.
- Wang, Y., Stingl, U., Anton-Erxleben, F., Zimmer, M., Brune, A., 2004. 'Candidatus *Hepaticola porcellionum*' gen. nov., sp. nov., a new, stalk-forming lineage of Rickettsiales colonizing the midgut glands of a terrestrial isopod. *Arch. Microbiol.* 181(4), 299-304.
- Wei, L., YuJuan, L., XiaoLiang, L., Ping Z., Hong, Y., 2016. *Clostridium perfringens* promotes the growth and development of *Drosophila melanogaster*. *Acta Entomol. Sin.* 59(5), 530-7.
- Wilson-Sanders, S.E., 2011. Invertebrate models for biomedical research, testing, and education. *ILAR J.* 52(2), 126-52.
- Zimmer, M., 2002. Nutrition in terrestrial isopods (Isopoda: Oniscidea): an evolutionary-ecological approach. *Biol. Rev. Camb. Philos. Soc.* 77(4), 455-93.