THE FIRST DETECTION OF Acuaria spinosa IN PHEASANTS

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Summary: Parasitoses caused by the *Acuariidae* helminths sporadically occur in both free-living and artificially bred pheasant populations (*Phasianus colchicus* L.). In order to verify this situation, an investigation with the aim of detecting the *Acuaria species*. infection on seven pheasant farms in Serbia and Romania, was carried out. A total of 127 adult birds were examined. The collected nematodes were identified by their morphometric characteristics. The three different *Acuaria* species were identified including: *Cheilospirura* (*Acuaria*) *hamulosa*, *Dyspharinx* (*Acuaria*) *spiralis* and *Acuaria spinosa*. The parasites identified as the *Acuaria spinosa* species had equal lips and furrow cuticula. Both parasites had four spiny cordons 'which did not exceed a third of the anterior of the esophagus. The tail was short and rounded. Eggs were ellipsoid, thick shelled, embryonated, 0.039-0.41 µm long by 0.025-0.027 µm wide.

Furthermore, thefirst parasite's body length was $32 \, \mu m$, with a width of $0.336 \, \mu m$ (maximal width at the middle of the body). Four spiny cordons were $0.797 \, \mu m$ long, and the cylindrical pharings were $0.018 \, \mu m$ long. The vulva lies just posterior to the middle of the body, $3.56 \, \mu m$ from the anterior end, $51 \, \%$ of the total body length (TBL) from the anterior end. The anus lies at $0.25 \, \mu m$ from the posterior end. The body length of the second parasite was $34 \, \mu m$ with a width of $0.367 \, \mu m$. Spiny cordons 'which did not exceed a third of the anterior of the oesophagus were $0.809 \, \mu m$ long, and the cylindrical pharings were $0.021 \, \mu m$ long. The vulva lies just posterior to the middle of the body, $5.64 \, \mu m$ from the anterior end, $57 \, \%$ of the total body length (TBL) from the anterior end. The anus lies at $0.28 \, \mu m$ from the posterior end.

According to the authors knowledge this is the first detection of the Acuaria spinosa in pheasants (Phasianus colchicus L.).

Key words: Acuariidae; Phasianus colchicus; Acuaria spinosa

Introduction

Parasitoses caused by nematodes produceshealth problems in both the free-living and artificially bred pheasant populations. The nematode infection is the most frequent infection transmitted through an intermediate host in the pheasant population. There are several papers regarding helminthoses in freeliving and farm bred pheasants. Most of these investigations were performed in Europe, namely

in Czechoslovakia, Lithuania, Poland, Greece, Germany and Italy (2, 8, 10, 12, 17, 25).

In the West Balkan countries, numerous investigations were conducted in Romania (5, 14) and Serbia (20, 22, 23, 24). During those examinations parasitoses caused by the *Acuariidae* helminthes occurred sporadically, in both free-living and farm bred populations of pheasants (9, 16, 29). Therefore, this investigation was carried out on seven pheasant farms in order to establish the helminth fauna with an emphasis on the presence of the *Acuaruidae* species in the artificially bred pheasant population in Serbia and Romania.

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Material and methods

The investigation was carried out on seven pheasant farms for hunting/restocking purposes including four farms in Serbia and three farms in Romania. The helminthes were collected from adult pheasants that had died of natural causes by parasitological necropsy. A total of 127 adult birds were examined. The collected nematodes were fixed in glacial acetic acid and preserved in 70 % ethanol. For further examination, parasites were clarified in lactophenol and nematodes were identified by their morphometric characteristics described by Skrjabin et al. (26) and Chabauda (4). All the laboratory techniques used were described by Šibalić and Cvetković (28).

Gizzards were dissected in a 0.85 % NaCl solution and the cuticle was removed for helminth investigation. Fragments of the parasitized gizzards were removed and immediately fixed in formalin. The material was then processed for paraffin embedding. Five micrometres thick sections were stained with hematoxylin and eosin.

Micrographs were obtained using a Carl Zeiss bright field microscope, by means of Practica PLC3 and a Kodachrome 64 film.

Results

Infection with nematodes was found in 37.79 % (48/127) cases. Polyparasitism involving two species was detected in 28.34 % (36/127) of pheasants (13,21).

A total of 14 of 127 (11 %) examined pheasants were found to have been infected with the Acuariidae sp. During necropsy, specimens were found lying freely under the gizzard cuticle, partially or fully burrowed in the walls of the organ, mainly in the region of the caudal blind sac. There, specific macroscopic lesions were observed as present in the muscular portion of the stomach and included sclerotic and nodular lesions accompanied by a fine-whitish granular residuum, located underneath the corneous lining of the gizzard, at the points of nematodes implantation (15). Three different species were collected and identified: Cheilospirura hamulosa, Dyspharinx spiralis, and Acuaria spinos, which was found in onlyt one bird examined, which was represented by two parasites. Furthermore, it was found that C. hamulosa and D. spiralis infections occurred in both countries (on one farm in Serbia and two in Romania), while A. spinosa infection occured only in Romania. *Acuaria spinosa* was found in only one bird, and two worms were found.

The parasites identified as the Acuaria spinosa species had equal lips and furrow cuticula. Both parasites had four spiny cordons 'which did not exceed a third of the anterior of the esophagus. The tail was short and rounded. Eggs were ellipsoid, thick shelled, embryonated, and 0.039-0.41 μm long by 0.025-0.027 μm wide.

Furthermore, the first parasite's body length was 32 μm , the width was 0.336 μm (maximal width at the middle of the body). Four spiny cordons were 0.797 μm long, and cylindrical pharings were 0.018 μm long (Fig. 1). The vulva lies just posterior to the middle of the body, 3.56 μm from the anterior end, 51 % of the total body length (TBL) from the anterior end. The anus lies at 0.25 μm from the posterior end.

The second parasite's body length was 34 μ m, and the width was 0.367 μ m. Spiny cordons did not exceed exceed a third of the anterior of the esophagus. Spiny cordons were 0.809mm long, and the cylindrical pharynx was 0.021 μ m long (Fig. 2). The vulva lies just posterior to the middle of the body, 5.64 μ m from the anterior end, 57 % of the total body length from the anterior end. The anus lies at 0.28mm from the posterior end.

The histological examination revealed traumatic lesions in hemorrhagic and necrotic routes and tunnels generated by cuticular cordons and spines of the adult nematodes, surrounded by a conjunctive reaction (Fig. 3). Nematodes were observed in tunnels with fibrous walls, impregnated with various mesenchimal cells, predominant lymphocytes and eosinophils. The muscular wall of the stomach was almost completely replaced by conjunctive tissue, in which diffuse lymphoid infiltration and lymphoid nodules, as well as parasitic granuloma were observed.

Discussion

Acuaria spinosa belongs to the Acuariidae family, the Acuariinae subfamily, and genus Acuaria. When first described in Bonasa umbellus, the species was named Cheilospirura spinosa (6, 7). In a later comparison withs nematode from the Acuariinea family it was concluded by Boughton (3) that they were the same species,

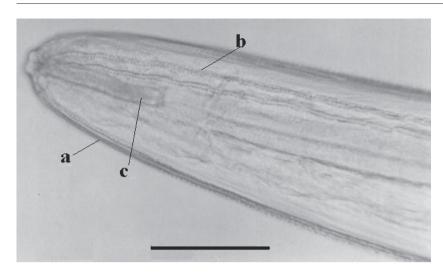


Figure 1: Anterior portion of a second sample of *Acuaria spinosa* ventral view, a) furrow cuticula, b) spiny cordons; c) pharings; Bar = 0.1 mm (Bar of 1 is common to 2 and 3)

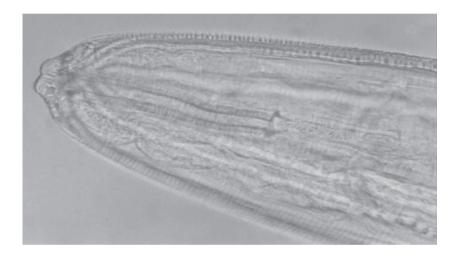


Figure 2: Anterior portion of a second sample of *Acuaria spinosa*, ventral view

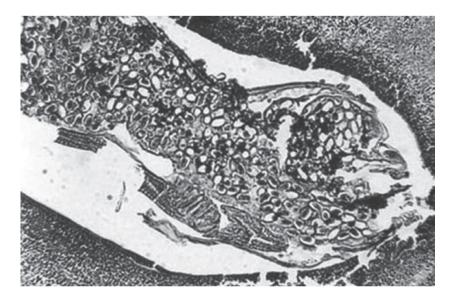


Figure 3: Perforated sites in the submucosa caused by *A. spinosa*

and that nematodes belong to the Acuaria genus. Finally, a complete description of parasites and their development was presented by Skrjabin et al. (25). According to Skrjabin et al., parasites measured 37-40 μm long and 0.31-0.34 μm wide and possessed two equal lips and furrow cuticula. These measurements coincide with the findings of this study (parasites body lengths were 32-34 μm and widths were 0.336- 0.367 μm ; four spiny cordons were 0.797- 0.809 μm long).

Grasshoppers (*Melanopus femurrubrum* and *M. differentialis*) serve as intermediate hosts for Acuaria spinosa. In these hosts the infective larvae develop and the final host acquire the infection by ingesting grasshoppers (7, 26).

Acuaria spinosa (syn. Cheilospirura spinosa) was first described as a species of nematode found in the gizzard of ruffed grouse in Michigan (USA) (27). Cram (6) detected A.spinosa in the horny lining of the gizzard of Bonasa umbellus, Colinus virginianus and Pediocetes (Tympanuchus) phasianellus campostris. Later, A. spinosa was found in Alectoris graeca (1) and in Tympanuchus cupido (9). The results of this study conclude that this is the first occurrence of Acuaria spinosa in ring necked pheasants (Phasianus colchicus L.).

The helminthes from the Acuariidae family sporadically occur in pheasants, as first reported in the USA by Cram (6). Later examination pointed to the factthat Acuariidae can be found in free living and farm bred pheasants in numerous countries in Europe (10, 11, 12, 13). During those examinations the following species were found: *C. hamulosa*, followed by *A. spiralis* and *A. gruveli*. In Serbia and Romania *C. hamulosa* was the most frequent Acuariida species (13, 14, 19, 20, 21).

In the infected birds, parasites of the Acuaridae genus caused severe gross lesions in the gizzard such as hemorrhages, ulcers and thickening of the mucosa and cuticle, and single yellowish nodules on the caudoventral muscle (16). Histological examination of the infected gizzards revealed discrete and coalescing nodular and cystic lesions in the mucosa and musculature which contained sections of the parasite. Cellular reaction in the lesions was characterised by a large number of lymphocytes, monocytes, plasma cells, heterophils, and, in some of the sections, a severe eosinophilic reaction. The mucosa and submucosa showed markedly thickened and diffuse mononuclear infiltration as well as reactive lymphoid nodules (15). During examinations it was found that pathological changes caused by *A. spinosa* were similar to cases of infection with other Acuaria species.

The presence of Acuaria spinosa in pheasants or other birds, except Bonasa umbellus, Colinus virginianus, Pediocetes (Tympanuchu) phasianellus, Alectoris graeca and Tympanuchus cupido was not reported in available literature. Furthermore, the presence of adult nematodes in gizzard suggests the possibility of adaptation of Acuaria spinosa to parasitism in a new host - pheasants. This is not unique, because Brachylaemus fuscatus and Plagorchis megalorchis were found in pheasants in Scotland (24). Zygocotyle lunata was found in 1.6 % of pheasants in Nebraska (11). Infection with Ornithostrongylus quadriradiatus in pheasants was found in the Belgrade area (18). During 2002 the first occurrence of Thominx cyanopicae was established in the new host - pheasants and their full adaptation to it, confirmed by histological aspects which revealed the pathological effect of parasites (14).

The first occurrence of *Acuaria spinosa* in pheasants has an epidemiological importance and indicates the necessity for continued parasitological examination of this bird species.

References

- 1. Akhumyan KS, Khanbegyan RA. The helmint fauna of wild galliforms in Armenia (*Coturnix coturnix, Alectoris graeca, Perdix perdix, Lyrulus mlokosiewiczi* and *Tetraogallus caspius*). Zool Sb Akad Nauk Armyanskoi SSR, Inst Zool 1982; 18: 9–45.
- 2. Arnasteikskene T, Kazlauskas Yu, Kadite B. Parasitic fauna of pheasants (*Phasianus colchicus* L.) in the Lithuanian SSR. Acta Parasitol Lith 1970; 10: 95–101.
- 3. Boughton B. *Cheilospirura spin*osa (Cram, 1927). Univ Minn Agr Exp Stat, 1937: 21–2. Techn Bull 121.
- 4. Chabauda G. Keys to the genera of the order Spirurida. Part 2. Spiruroidea, Habronematoidea and Acuarioidea. In: Anderson RC, Chabaud Willmott AS, eds. CIH keys to the nematode parasites of vertebrates. No. 3. Farnham Royal: Commonwealth Agricultural Bureaux,, 1975: 29–58.
- 5. Cosoroabă J, Ciolofan I. Controlul capilariozei si singamozei in crescatoriile de fazani si potârnichi. Lucr Sti Med Vet 1985; 27: 69–73.
 - 6. Cram EB. Bird parasites of the nematodes

suborders Strongylata, Ascaridata and Spirurata. US Nat Mus Bull 1927; 140: 334–61.

- 7. Cram EB. Developmental stages of some Nematodes of the Spiruroidea parasitic in poultry and game birds. US Dept Agriculture, Tech Bull 1931: 227–8.
- 8. Gassal S, Schmäschke R. The helminth and coccidial fauna of pheasants (*Phasianus 1colchicus*) in view of the specific environmental conditions in pheasantries and in the wild. Berl Munch Tierarztl Wochenschr 2006; 119: 295–302.
- 9. Gibson DI, Bray RA, Harris EA (Compilers) Host-parasite database of the Natural history museum, London 2005. http://www.nhm.ac.uk/research-curation/scientific-resources/taxonomy-systematics/host-parasites/index.html (Dec. 2013)
- 10.Githkopoulos PR. Helminths in pheasants of Greek phesanteries. Hellenike Kten 1984; 27: 68–76.
- 11. Greiner EC. Parasites of Nebraska pheasants, 63 Phasianus colchicus. J Wildl Dis 1972; 8: 43–5.
- 12. Hospes R. Parasitosen des Jagdfasans. PhD dissertation. Giessen: University of Giessen, 1996.
- 13. Floristean I. Examination of etiology, morphopathology and clinic of endoparasites of pheasants artificially raised. PhD dissertation. Iasi, Romania: University of Iasi, 2003: 225 str.
- 14. Floristean I, Pavlović I. The first occurence of *Thominx cyanopicae* (Lopez Neyra, 1947) in pheasants (*Phasianus colchicus* L.). Acta Vet Beograd 2003; 53: 183–90.
- 15. Floristean I, Cotofan O, Merticariu S, Pavlović I. Etiological and morphopathological investigations in acuariosis in breeding pheasants. Lucr Sti Med Vet 2003; 46:141–4.
- 16. Menezes RC, Tortelly R, Gomes DC, Pinto RM. Pathology and frequency of *Cheilospirura hamulosa* (Nematoda, Acuarioidea) in Galliformes hosts from backyard flocks. Avian Pathol 2003; 32:151–6.
- 17. Okulewitz A, Modrezejavska M. Helminth fauna of pheasants (*Phasianus colchicus* L.) from the environs of Wroclaw in the autumn/summer period. Wiad Parazitol 1980; 26: 73–5.
- 18. Pavlović I, Hudina V, Kerš V, Blažin V, Čupić V. Helminthological fauna of pheasants in Belgrade zoo in period March-August 1988. Vet Glasn 1990; 44: 467–71.
- 19. Pavlović I. Ecto and endoparasites of pheasants artificially raised and measure to its control. MSc thesis. Belgrade, Serbia: University

- of Belgrade, 1991: 76 str.
- 20. Pavlović I, Kerš-Pavlović V, Jordanović B, Hudina V. Endoparasites of pheasants artificially raised. Lucr Sti Med Vet 1992; 26:104–7.
- 21. Pavlović I, Jakić-Dimić D, Kulišić Z, Florestean I. Most frequent nematode parasites of artificially raised pheasants (*Phasianus colchicus* L.) and measures for their control. Acta Vet Beograd 2003; 53: 393-8.
- 22. Pavlović I, Dačić M, Stokić-Nikolić S, Rajković M, Ivanović S, Đorđević M. Nematode parasites of pheasants (*Phasianus colchicus* L.) artificially raised in Serbia. Lucr Sti Ser D Zoot Anim Sci 2009; 52: 412–15.
- 23. Pavlović I, Đorđević M, Kulišić Z. Endoparasites of farm-reared pheasants (*Phasianus colchicus* L.) in Serbia. In: Modern aspects of sustainable management of game population. International symposium on hunting, Zemun-Belgrade, Serbia, 2012: 125–9.
- 24. Rayski C. An outbreak of helminthosis in pheasants chicks due to *Plagiorchis megalorchis* (Ress, 1952) with same critical remarks on P. (M) larcola Skrjabin 1924. Parasitology 1964; 54: 391–6.
- 25. Santilli F, Bagliacca M. Occurrence of eggs and oocysts of intestinal parasites of pheasant (Phasianus colchicus) in Tuscany (Italy) Eur J Wildlife Res 2012; 58: 369–72.
- 26. Skrjabin, KI, Soboleva NDV, Ivashkin M. Principles of nematology Spirurata of animals and man and the diseases caused by them. Part 3. Moscow: Izdatel'stvo Academii Nauk SSSR,1965: 131–2.
- 27. Stafseth HJ, Kotlan A. Report of investigations on an alleged epizootic of ruffed grouse in Michigan-America. J am Vet Med Assoc 1925; 67: 260–7.
- 28. Šibalić S, Cvetković Lj. Osnovi dijagnostike parazitskih bolesti Beograd: OZID, 1980: 36–7
- 29. Tampieri MP, Galuppi R, Rugna G. Survey on helminthofauna in pheasants from Eastern Europe. Parassitologia 2005; 47: 241–5.

PRVO ODKRITJE Acuaria spinosa PRI FAZANIH

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Povzetek: Parazitoze, ki jih povzročajo črevesne gliste iz družine Acuariidae, naj bi se občasno pojavljale pri prosto živečih in gojenih populacijah fazanov. Opravili smo raziskavo, s katero smo želeli odkriti okužbo z Acuaria species na sedmih farmah fazanov v Srbiji in Romuniji. Skupno je bilo preiskanih 127 odraslih ptic. Zbrani nematodi so bili določeni glede na njihove morfološke lastnosti. Identificirali smo tri različne vrste Acuaria: Cheilospirura (Acuaria) hamulosa, Dyspharinx (Acuaria) spiralis in Acuaria spinosa. Zajedavci, ki so bili identificirani kot Acuaria spinosa, so imeli enake ustnice in nagubano kutikulo. Obe vrsti zajedavca sta imeli po 4 trnaste kordone, ki niso presegali tretjine sprednjega dela požiralnika. Njihov rep je bil kratek in zaokrožen. Jajčeca so bila embrionirana, elipsoidne oblike z debelo lupino, dolga 0,039-0,41 µm in široka 0,025-0,027 µm.

Telesna dolžina prvega zajedavca je bila 32 µm s širino 0,336 µm (maksimalna širina na sredini telesa). Štirje trnasti kordoni so bili dolgi po 0,797 µm, valjasto žrelo pa 0,018 µm. Vulva je ležala posteriorno na sredini telesa, oddaljena 3,56 µm od sprednjega konca in 51-odstotkov skupne dolžine telesa od anteriornega dela. Anus je ležal 0,25 µm od posteriornega dela. Telesna dolžina drugega parazita je bila 34 µm, širina pa 0,367 µm. Trnasti kordoni ki niso presegli tretjine anteriornega dela požiralnika, so bili dolqi 0.809 µm, valjasto žrelo pa 0.021 µm. Vulva je ležala posteriorno od sredine telesa, oddaljena 5.64 µm od anteriornega konca in 57-odstotkov skupne dolžine telesa od anteriornega dela. Anus je ležal 0,28 µm od posteriornega konca.

Po naših podatkih je to prvi primer odkritja Acuaria spinosa pri fazanih (Phasianus colchicus L.).

Ključne besede: Acuariidae; Phasianus colchicus; Acuaria spinosa