

Agrovoc descriptors: nematoda, life cycle, slugs, mortality, parasites, identification, diagnosis, monitoring

Agris category code: H10

Massive occurrence and identification of the nematode *Alloionema appendiculatum* Schneider (Rhabditida: Alloionematidae) found in Arionidae slugs in Slovenia

Žiga LAZNIK¹, Jenna L. ROSS², Stanislav TRDAN³

Received October 26, 2009; accepted January 18, 2010.

Delo je prispelo 26. oktobra 2009; sprejeto 18. januarja 2010.

ABSTRACT

In the period from June to October 2008 we collected 500 slugs from the genus *Arion* in the area of Ljubljana and Prekmurje (Slovenia). By means of dissection we determined the presence of parasitic nematodes in slug cadavers. Identification of the nematodes was made by a molecular technique (PCR). In these slugs we did not find the parasitic nematode *Phasmarhabditis hermaphrodita*, however the presence of *Alloionema appendiculatum* in larger quantities was confirmed. The most infected was a Spanish slug, *Arion lusitanicus*. In Petri dishes younger slugs showed a satisfactory mortality rate already on the fourth day after the application of the nematode suspension. Unfortunately, we can not confirm with certainty that the nematode *A. appendiculatum* undergoes a complete life cycle in *A. lusitanicus*, which is otherwise typical for *Phasmarhabditis hermaphrodita*.

Key words: *Alloionema appendiculatum*, *Arion lusitanicus*, parasitic nematodes

IZVLEČEK

ŠTEVILČNI POJAV IN IDENTIFIKACIJA OGORČICE *Alloionema appendiculatum* Schneider (Rhabditida: Alloionematidae) V LAZARJIH (Arionidae) V SLOVENIJI

V obdobju od junija do oktobra 2008 smo na območju Ljubljane in Prekmurja nabrali 500 polžev iz rodu *Arion*. Polže smo secirali in ugotavljali zastopanost ogorčic v njihovem telesu. Identifikacija ogorčic je bila opravljena z molekulsko tehniko (PCR). V nobenem polžu nismo našli parazitske ogorčice *Phasmarhabditis hermaphrodita*, medtem ko smo zastopanost ogorčice *Alloionema appendiculatum*, predvsem v predstavnikih vrste *Arion lusitanicus*, potrdili v večjem številu. V petrijevkah so mladi polži pokazali zadovoljivo stopnjo smrtnosti že četrty dan po nanosu suspenzije ogorčic. Žal pa z našo raziskavo ne moremo potrditi, da ogorčica *A. appendiculatum* v polžu *A. lusitanicus* razvije popolni parazitski razvojni krog, kot je to značilno za ogorčico *P. hermaphrodita*.

Ključne besede: *Alloionema appendiculatum*, *Arion lusitanicus*, parazitske ogorčice polžev

1 INTRODUCTION

More than 25,000 species of nematodes have been described up to the present, of which around 4,000 are marine free-living nematodes, 6,000 terrestrial free-living nematodes, 12,000 parasitic nematodes of vertebrates, and 3,500 parasitic nematodes of invertebrates (Poulin and Morand, 2000; Hugot *et al.*, 2001). Hitherto, in terrestrial slugs nematodes from eight families have been determined. Among them are parasitic species (of the order Rhabditida) which spend

their complete life cycle inside slugs, as well as nematodes (of the order Strongylida) which use slugs as intermediate hosts (Grewal *et al.*, 2003).

When compared with entomopathogenic nematodes - the natural enemies of insects (Kaya and Gaugler, 1993) - these nematodes, which are related to slugs, are poorly studied (Wilson and Grewal, 2005). However, among those which have been studied up to now, specificity to

¹ Young researcher, B. Sc., Jamnikarjeva 101, SI-1111 Ljubljana, email: ziga.laznik@bf.uni-lj.si

² Ph. D, University of Aberdeen, Institute of Biological and Environmental Sciences, AB24 3UU, UK

³ Assoc. Prof., Ph. D, Jamnikarjeva 101, SI-1111 Ljubljana

host species as well as diversity among individual species of nematodes are known. Slug-parasitic nematodes were found in all organs of the body cavity and also on foot muscles. Infection mechanisms are different, from entering through the body wall (infective juveniles of nematodes which are found in the soil) to the transference of infective juveniles between slugs during mating. It is well known that slug-parasitic nematodes can be an important biotic factor which balances the slug population in the natural environment (Grewal *et al.*, 2003). The most intensively studied species of slug-parasitic nematode is *Phasmarhabditis hermaphrodita* (Schneider) (Rhabditidae), and in some countries this species is also used in the biological control of slugs (Wilson *et al.*, 1993ab). The natural hosts of *P. hermaphrodita* are numerous slug species which can also be found in Slovenia: *Deroceras leave* (Müller), *D. reticulatum* (Müller), *Arion distinctus* Mabille, *A. lusitanicus* Mabille, *A. silvaticus* Lohmander, *Tandonia budapestensis* (Hazay) and *T. sowerbyi* (de Férussac) (Vaupotič and Velkovrh, 2002; Morand *et al.*, 2004).

Alloionema appendiculatum Schneider is classified into the family Alloionematidae and it consists of two genera, *Rhabitophanes* Fuchs, the species of which are most frequently associated with insects, and the genus *Alloionema* Schneider, whose only representative, *A. appendiculatum*, is linked with gastropod molluscs (Morand *et al.*, 2004). The former species was discovered for the first time in 1859 (Schneider). Its presence has hitherto been confirmed in Europe and Australia (Cabaret *et al.*, 1988). In Slovenia this species was confirmed as early as in the 1970s, when Hržič studied the nematological fauna of Slovenia. The nematode was found in the soils of a vineyard situated in the Primorska region of Slovenia (Hržič, 1969; Urek *et al.*, 2003). With our research we also wanted to determine if in Slovenia this nematode species is also present in slugs, as they are becoming increasingly more important pest organisms of some cultivated plants (Barker, 2001).

2 MATERIAL AND METHODS

In September and October 2008 we collected 500 slugs in the area of Prekmurje and Ljubljana from the genus *Arion* (Arionidae). We dissected the slugs and with electronic magnification searched for the possible presence of nematodes inside them. We confirmed the presence of nematodes in the slugs collected in Ljubljana. With a pipette we separated the nematodes from the slug tissue and cleaned them according to the prescribed protocol (Riddle 1988). We infected new slugs with the obtained nematode suspension. We put 1 ml of a nematode suspension with a concentration of 300 IJ/ml into a Petri dish (diameter = 9 cm) and added 5 smaller slugs of the species *Arion lusitanicus* Mabille. After 4 days the slugs died and we dried the dead individuals at air temperature for 10 days. On the 10th day we placed the dried cadavers into a so-called White trap (Bedding and Akhurst 1975) in order to separate the nematodes from the dead slugs. The nematodes started to leave the cadavers on the 4th day after being exposed to the White trap. We washed the nematodes according to a procedure using a centrifuge and a 5 % concentration of sodium hypochlorate. The aim of this process was to obtain infective juveniles from the suspension. The nematode suspension was sent to the Institute of Biological and Environmental Sciences, University of Aberdeen, for further analysis.

DNA was extracted using a QIAmp DNA Mini Kit. The 18S gene was amplified using a combination of universal primer pairs: G18S4, 26R, 24F, and 18P (Blaxter *et al.* 1998). An additional primer pair, 22F, and a newly designed primer, 1080R (TCC-TGG-TGG-TGC-CCT-TCC-GTC-AAT-TTC), were also used to ensure complete coverage of the 18S region (J. Ross, unpubl.). PCR cycling parameters involved primary denaturation at 94°C for 5 minutes, followed by 35 cycles of 94°C for 60 s, 55°C for 90 s, and 72°C for 2 min. Post-amplification extension occurred at 72°C for 10 min (J. Ross, unpubl.). The PCR products were visualized on a 1% agarose gel and cleaned using a Qiagen QIAquick® PCR Purification Kit (fig. 1). Sequencing was carried out using an ABI3730 sequencer. The sequences were assembled using Sequencer 4.1 (Genes Codes Corp. Ann Arbor, Michigan, USA). The results were then compared with the GenBank Database, (<http://www.ncbi.nlm.nih.gov/>), using the BLASTn search tool (Altschul *et al.* 1990). The *Alloionema appendiculatum* sequence was deposited in the NCBI GenBank under FJ665982.

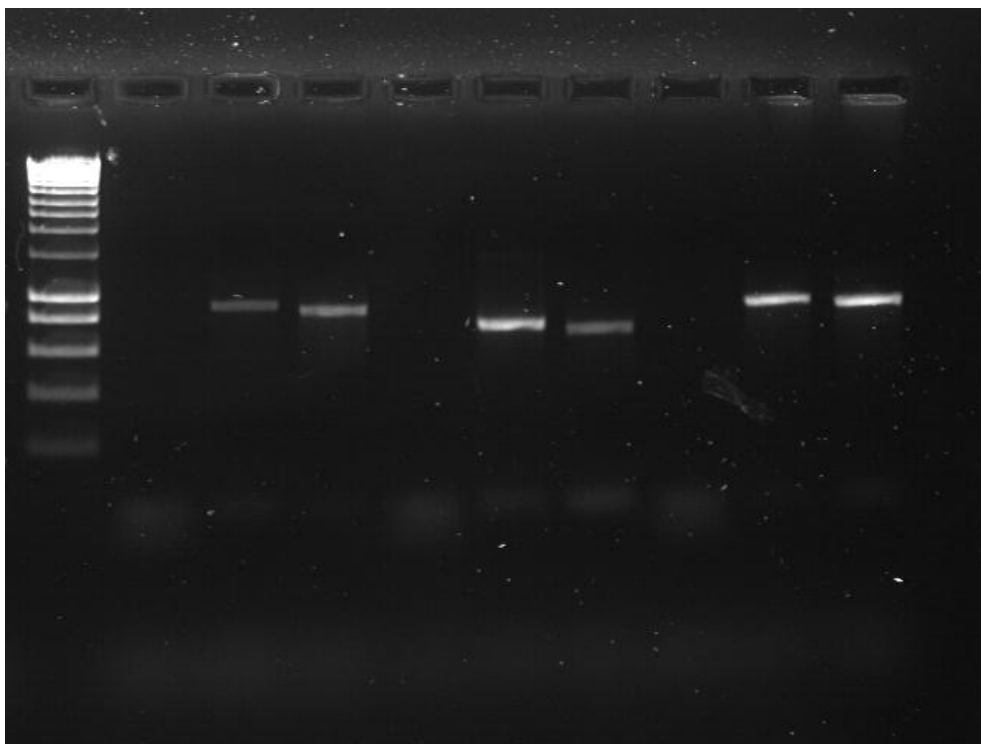


Figure 1: 1% TAE buffered agarose gel, in the 1st lane; GeneRuler 100 bp DNA Hyperladder (Fermentas), in the 3rd, 6th, and 9th lane; the PCR product of *Phasmarhabditis hermaphrodita* (Becker Underwood commercial strain), which was used as a positive control, using the primer pair specified in the text; the 4th, 7th and 10th lane: the PCR product of the sample slug nematode – *Alloionema appendiculatum*.

3 RESULTS AND DISCUSSION

The genetic studies proved that this nematode species is *Alloionema appendiculatum* Schneider (1859). The closest matches to this *Alloionema appendiculatum* sequence were *Rhabditoid* sp. (DQ531722.1) (Qvarnstrom *et al.*, 2007) with 92% identity over 89% coverage, followed by *Strongyloides ratti* (U81581.1) (Frisse *et al.*, unpubl.) with 87% identity over 100%

coverage (Fig. 2). The ITS1-5.8S-ITS2 region, including the partial 18S and 28S rRNA genes (flanked by the above primers) of the Slovenian isolate of *A. appendiculatum* is 1425 bp long (fig. 2).

FJ665982	1	AACTGAGGTAATTCTTGAGCTAATACACGC--T--T--TA-ATGCCACATT-CGTGGTGC	52
U81581	86	T-...A.....-A.....	138
FJ665982	53	GTTTATTTGACTAGA-C-A---T-C-A-TA-T-T-GGTTGACTCAAAATATCCTTGCT-A	100
U81581	139	T..A.C-...-T-.T-.-.C.....C...-G	187
FJ665982	101	ATTTTTTTTTTA-AAAAT-ATGCCGTATGAGTATCTGCTTTATCAACTTTTCGATGGTAGG	158
DQ531722	1		6
U81581	188	G..AC--...C-..A.....T.....G.....	244
FJ665982	159	GTATTGGCCTACCATGG-TGTTGACGGATAACGGAGAATTAGGGTTCGACTCCGGAGAGG	217
DQ531722	7	-A.....	65
U81581	245	T..-.....	303
FJ665982	218	GAGCCTGAGAAACGGCTACCACATCCAAGGAAGGCAGCAGGCGCGAAAATTACCCAATCT	277
DQ531722	66	T.	125
U81581	304	T.	363

FJ665982	278	TAG-TT-AAAGGAGGTAGTGACGAAAAATGACAAAG-C-AA-CACT-TT-TTAAG-T-GT	328
DQ531722	126	...-...-...A.....C...T-G...-...T...-A.	176
U81581	364	...-...-...A.....C...A...T.T.A...T...AT.	416
FJ665982	329	TGTTATTGGAAATCTTGC-AA-CCTAAAT-A-GTTGCTTGGTAAAGGAGAGGGCCAGTCT	384
DQ531722	177	G.....T...G-TG.....-AC.A.....	232
U81581	417	...G...A.....GTT....A.-C.....A....A....	471
FJ665982	385	GGTGCCAGCAGCCGCGGTAATACCAGCTCTCCAAGTGCATAAAATGATTGTTGTGGTTAA	444
DQ531722	233	T.....	292
U81581	472	T.....	531
FJ665982	445	AAAGCTCGTA-GTTGGATT-T----ATTG---AATA-A-G--T-TCA-GA-G---T-C--	482
DQ531722	293	-.....-.....G...G...-...-T...T--G--	331
U81581	532	-.....-.....A---G.TT---G.A.T...CA---	572
FJ665982	483	--A-T-GT-A-T-ATG-GCT--T--C--TGA-A-AT-G-TT---CA-TA-C--T-TT--T	514
DQ531722	332	--.C...-G...-A.A.A--...-...C...C..AAAT.T...G--A...-	369
U81581	573	--T...G...GT.A-TT...-A..T.CC...C...-...T-T--...-	606
FJ665982	515	-G-C---A-A---TT-AG---C-A-AT-A-A-T--T-GC-CT-GTTACTTTGAATAAATC	551
DQ531722	370	-T-T---T-T---.A.A---G...A...-A--A...-A...-	408
U81581	607	AT-T---.T...-...-A...A.T...AA.-AA...-...	649
FJ665982	552	AGAGTGTTCAAA--A--CA-G---TC-A---T-ATGAC-TG-A--AT-A-T-CCTAGCAT	591
DQ531722	409	-.....-.....-.....-.....	448
U81581	650	...G.....-C---G.CA-.T...-...T...-T...GG-.....	692
FJ665982	592	GGAATAACACTATA-GAAGTATATG-TAGTGCGGTGTTTCA-C--ACA-T-TACTTCATG	644
DQ531722	449	-.....-T.....T.....TT--...-A.....	500
U81581	693	-.....-A.T.....T...-...-T...C.TT.....	741
FJ665982	645	GTTAATA-GAAACAAACGGGGGCATTTCGTATCGATACGTTAGAGGTGAAATTCCTTGGACC	703
DQ531722	501	-.....	559
U81581	742	A.....-G.....C.....	800
FJ665982	704	GTATCGAGACGTCCTACTGCGAAAGCATTGCGCAAGAATGTTTTTCAATCAAGAACGA	763
DQ531722	560		619
U81581	801	...G.....	860
FJ665982	764	AAGTTAGAGGTTTGAAGGCGATCAGATACCGCCCTAGTTCTAACCGTAAACTATGCCTAC	823
DQ531722	620	G.....	679
U81581	861		920
FJ665982	824	TAGATGTTTG-A-TGGTA-AT-GTA-AAT-TA-TTA-TTGA-GCATCTTCTCGGAAACGA	874
DQ531722	680	A...-A...-...-...-...T.....	730
U81581	921	...G...A..A...-...-T.A...-T...A...A...T.T...C.....	971
FJ665982	875	AAGTCTTTCGTTCCGGGGGAAGTATGGTTGCAAAGCTGAAACTTAAAGGAATTGACGGA	934
DQ531722	731		790
U81581	972		1031
FJ665982	935	AGGGCACCACCAGGAGTGGAGCCTGCGGCTTAATTTGACTCAACACGGGAAAACCTCACCC	994
DQ531722	791		850
U81581	1032	N.....	1091
FJ665982	995	GGGCCGGACA-TAAG-AAGGATTGACAGATTTAGAGCTCTTTCAAGATTTTA-TGGTTGG	1051
DQ531722	851	-CG.T-.....AA.....T.....TT.....CG.....	907
U81581	1092	C..T--.....G.T.....T...-G.....	1148
FJ665982	1052	TGGTGCATGGCCGTTCTTAGTTCGTGGATATGATTTGTCTGGTTGATTCCGATAACGAAC	1111
DQ531722	908		967
U81581	1149	G.	1208

FJ665982	1112	GAGACTGTAA-T-T-TAT-T---T---T-AC--AC-G-TGGGGG-ATTC-T-T--A-A-	1149
DQ531722	968	C.....-A-----G--T-A-.ACATT-...TA.-.-G-.-	1006
U81581	1209	T....-G....-A-----A--T---.ATATT-...T-.-T-.-	1246
FJ665982	1150	-TCC---T--T-TA-ACGGTT--AATAA-ATGATTAGT-GA-TGTTTAA-TCACCTTGAGA	1195
DQ531722	1007	-.AG---.-GA..T-----T-----GC.-...G-.....	1052
U81581	1247	-.AA---A-.-...TT-----C-----A.A.--.....C.-T.....	1292
FJ665982	1196	AAGAGCAATAACAGGTCTGTGATGCCCTTAGATGTCTGGGGCTGCACGCGCTACAATG	1255
DQ531722	1053	.G.....	1112
U81581	1293	G.....G.....A.....C.....	1352
FJ665982	1256	TAGTGAGCA-CTATGTTCTGATTC-GAGAGGATTGGGTAAACCTTGAAAGCATTACGTA	1313
DQ531722	1113	T.-.....	1170
U81581	1353	T.-T.....T..A-....TA.A.....A.....A.....	1410
FJ665982	1314	ACTGGGAGTGTAACCTTGCAATTATGTTACATGAACGAGGAATTCCAAGTAAACGTGAA-T	1372
DQ531722	1171	C....-	1229
U81581	1411	A.....T.....-..G.	1469
FJ665982	1373	CATTAG-TTCACATTGATTACGTCCCTGCCCTTTGTACACACCGCCGCTCGCTG	1425
DQ531722	1230	-...G.....	1282
U81581	1470	C.-.....	1522

Figure 2: Multiple sequence alignment of the ITS rDNA region (including partial fragments of the 18S and 28S rRNA genes) of 3 nematode species. Code FJ665982 corresponds to the Slovenian isolate of *Alloionema appendiculatum*. Codes DQ531722 and U81581 refer to the Rhabditoid sp. and *Strongyloides ratti*, respectively.

4 CONCLUSIONS

For the nematode *Alloionema appendiculatum* it is acknowledge that it has a free-living and parasitic life cycle (Cabaret and Morand, 1990). In the parasitic part, the larvae of the third larval stage (L3) enter into the slug's body through its foot, where they moult and undergo a fourth larval stage (L4) and which stays encapsulated in foot muscles. The encysted larvae engorge (mechanism unknown) and then exit the slug and molt into immature adults (L5), which are free-living (Morand and Daguzan, 1986). Adult nematodes which undergo a series of molts in the parasitic life cycle are two to three times longer than those (from 1 to 2 mm) which grow in a free-living life cycle.

The nematode *A. appendiculatum* has hitherto been found in different species of slugs; *Deroceras* sp. (Schneider, 1859), *Arion ater* (Schuurmans – Stekhoven, 1950), *A. circumscriptus* (Mengert, 1953), *A. intermedius* (Cabaret *et al.*, 1988), *A. silvaticus* (Morand and Bonnet, 1989), *A. subfuscus* (Cabaret and Morand, 1990), *Cantareus aspersus* (Charwat and Davies 1999), and *Prietocella barbara* (Morand, 2004). Research studies have not yet found a natural host in which the nematode *A. appendiculatum* undergoes a complete life cycle and which causes it to have a higher mortality rate (Morand *et al.*, 2004). Morand and Daguzan (1986) report that the nematode *A. appendiculatum* does not appear very frequently in

natural slug populations, meanwhile its presence in snails which have been bred is more common (Cabaret *et al.*, 1988).

A research study which was conducted by Cabaret and Morand (1990) indicates that a positive interaction exists between the nematodes *Muellerius capillaris* (Strongylida: Strongyloididae) and *A. appendiculatum*, as with the species *M. capillaris* previously infected slugs attain a higher mortality rate when infected with *A. appendiculatum* afterwards. The application of *A. appendiculatum* in practice might be carried out in soils where a larger number of nematodes from the family Strongyloididae is present and for which it is known that slugs are only intermediate hosts (Grewal *et al.*, 2003). But for now, no such co-infection has been discovered (Morand *et al.*, 2004).

This is the first large-scale record of the mentioned nematode in slugs from the genus *Arion*, among which *Arion lusitanicus* Mabile predominated in terms of the number of individuals. Younger slugs of the mentioned species showed a satisfactory mortality rate already on the fourth day of the nematode suspension application in Petri dishes. But because these results are merely preliminary, we can not yet confirm for certain that the nematode *A. appendiculatum* is capable of developing a complete life cycle inside the slug *A. lusitanicus*,

although this is common for *Phasmarhabditis hermaphrodita* (Wilson *et al.*, 1993a). The presence of *P. hermaphrodita* has not yet been confirmed in Slovenia, thus only chemical plant protection products with limacide activity can be used to control slugs and snails (Ozimič *et al.*, 2007).

Due to the fact that the Regulations on Biological Plant Protection prohibit the introduction of exotic organisms into the natural environment, we will continue our

research on the presence of slug-parasitic nematodes. Possible future identification of the nematode *P. hermaphrodita* could entail an alternative to chemical limacides, as due to the milder winters and humid summers, slugs and snails are increasingly important pests on cultivated plants in Slovenia as well as in many other countries (Kozowski *et al.*, 2006; Grubišić *et al.*, 2007).

5 ACKNOWLEDGEMENT

This work was carried out within Horticulture No P4-0013-0481, a program funded by the Slovenian Research Agency, and within the L4-1013 project funded by the Slovenian Research Agency and Ministry of Agriculture, Food, and Forestry of the Republic of Slovenia and the company Unichem d.o.o. Part of the

research was funded within Professional Tasks from the Field of Plant Protection, a program funded by the Ministry of Agriculture, Forestry, and Food of Phytosanitary Administration of the Republic of Slovenia. This study was also supported by Becker Underwood and BBSRC.

5 REFERENCES

- Altschul, S.F., Gish, W., Miller, W., Myers, E.X., Lipman, D.J. 1990. Basic local alignment search tool. *J. Mol. Biol.* 215: 403-410.
- Barker, G.M. 2001. The biology of terrestrial gastropods. CAB Int., Wallingford: 468 pp.
- Bedding, R. A., Akhurst, R. J. 1975. Simple technique for the detection of insect parasitic rhabditid nematodes in soil. *Nematologica* 21: 109-110.
- Blaxter, M.L., De Ley, P., Garey, J. 1998. A molecular evolutionary framework for the phylum Nematoda. *Nature* 392: 71-75.
- Cabaret, J., Morand, S., Aubert, C., Yvoré, P. 1988. Snail farming: a survey of breeding management, hygiene and parasitism of the garden snail, *Helix aspersa* Müller. *J. Mollus. Stud.* 54: 209-214.
- Cabaret, J., Morand, S. 1990. Single and dual infection of the land snail *Helix aspersa* with *Muellerius capillaris* and *Alloionema appendiculatum* (Nematoda). *J. Parasitol.* 76: 579-580.
- Charwat, S.M., Davies, K.A. 1998. A nematode isolate for biocontrol of pest snails in south Australia. *Nematologica* 44: 469-470.
- Grewal, S.K., Grewal, P.S., Hammond, R.B. 2003. Susceptibility of North American native and non-native slugs (Mollusca: Gastropoda) to *Phasmarhabditis hermaphrodita* (Nematoda: Rhabditidae). *Biocontrol Sci. Technol.* 13: 119-125.
- Grubišić, D., Gotlin-Čuljak, T., Vitas, I. 2007. Puževi golači u ratarskim kulturama - procjena rizika i mjere suzbijanja. *Glasnik biljne zaštite* 7: 394-398.
- Hržič, A. 1969. Investigation of nematodes as direct plant pests and vectors of plant diseases. Agricultural Institute of Slovenia, Ljubljana, Technical Report, 26 pp.
- Hugot, J.P., Baujard, P., Morand, S. 2001. Biodiversity in helminths and nematodes as a field of study: an overview. *Nematology* 3: 1-10.
- Mengert, H. 1953. Nematoden und Schnecken. *Z. Morphol. Okol. Tiere* 4: 311-349.
- Kaya H.K., Gaugler R. 1993. Entomopathogenic nematodes. *Ann. Rev. Entomol.* 38: 181-206.
- Kozowski, J., Zielinska, M., Pawowska, A., Kozowska, M. 2006. Susceptibility of some vegetable species to feeding of *Cepaea hortensis* (Muller) and *Arion rufus* (Linnaeus). *J. Plant Prot. Res.* 46: 231-239.
- Morand, S., Bonnet, J.C. 1989. Importance des nématodes en héliiculture et méthode de prophylaxie. *Haliotis* 19: 69-75.
- Morand, S., Daguzan, J. 1986. Contribution à l'escargot petit-gris (*Helix aspersa* Müller): Premiers résultats concernant l'acarien *Riccardoella limacum* (Schrunk) et le nématode *Alloionema appendiculatum* (Schneider). *Haliotis* 15: 31-39.
- Morand, S., Wilson, M.J., Glen, D.M. 2004. Nematodes (Nematoda) Parasitic in Terrestrial Gastropods. In Barker, G.M. (ed.) *Natural Enemies of Terrestrial Molluscs*. CABI Publishing, Wallingford, UK, 644 pp.
- Poulin, R., Morand, S. 2000. The diversity of parasites. *Q. Rev. Biol.* 75: 277-293.
- Riddle, D.L. 1988. The dauer larva, p. 393-412. In W.B. Wood (ed.), *The nematode Caenorhabditis elegans*, Cold

- Spring Harbor Laboratory Press, Plainview, New York, NY.
- Qvarnstrom, Y., Sullivan, J.J., Bishop, H.S., Hollingsworth, R., Da Silva, A.J. 2007. PCR-Based Detection of *Angiostrongylus cantonensis* in Tissue and Mucus Secretions from Molluscan Hosts. *Appl. Environ. Microbiol.* 73: 1415-1419.
- Schneider, A. 1859. Monographie der Nematoden. Druck und Verlag Von Georg Reimer, Berlin.
- Schuurams-Stekhoven, L.H. 1950 *Alloionema appendiculatum*. *Acta Zool. Lilloana* 9: 481-485.
- Urek, G., Širca, S., Karssen, G. 2003. A review of plant-parasitic and soil nematodes in Slovenia. *Nematology* 5: 391-403.
- Vaupotič, M., Velkovrh, F. 2002. Slugs (Gastropoda: Pulmonata: Milacidae, Limacidae, Boettgerillidae, Agriolimacidae, Arionidae) of Slovenia. *Acta Biol. Slov.* 45: 35-52.
- Wilson, M.J., Grewal, P.S. 2005. Biology, Production and Formulation of Slug-parasitic Nematodes. In Grewal, P.S., Ehlers, R.-U. And Shapiro-Ilan, D.I. (eds.) *Nematodes as biocontrol agents*. CABI Publishing, Wallingford, UK, 505 pp.
- Wilson, M.J., Glen, D.M., George, S.K. 1993a. The rhabditid nematode *Phasmarhabditis hermaphrodita* as a potential biological-control agent for slugs. *Biocontrol Sci. Technol.* 3: 503-511.
- Wilson, M.J., Glen, D.M., George, S.K., Butler, R.C. 1993b. Mass cultivation and storage of the rhabditid nematode *Phasmarhabditis hermaphrodita*, a biocontrol agent for slugs. *Biocontrol Sci. Technol.* 3: 513-521.