

# Influence of Ag nanoparticles on physiological and biochemical aspects of callus of *Thymus* species and *Zataria multiflora* Boiss.

Nima MOSAVAT<sup>1,2,3</sup>, Maryam YOUSEFIFARD<sup>4</sup>, Pooran GOLKAR<sup>5,6</sup>, Rabia JAVED<sup>7</sup>

Received September 12, 2020; accepted August 31, 2022.  
Delo je prispelo 12. septembra 2020, sprejeto 31. avgusta 2022

**Influence of Ag nanoparticles on physiological and biochemical aspects of callus of *Thymus* species and *Zataria multiflora* Boiss.**

**Abstract:** *Thymus* species have found remarkable importance in food and medicine industries. The present study investigates the potential effect of Ag nanoparticle elicitors on proliferation of callus, and production of carvacrol and thymol in *Zataria multiflora* and three *Thymus* species. Firstly, callus was induced on Murashige and Skoog (MS) medium containing 2 mg l<sup>-1</sup> of 2, 4-dichlorophenoxy acetic acid (2,4-D) and 1 mg l<sup>-1</sup> of kinetin (Kin)). Secondly, the effects of two different concentrations of Ag nanoparticles (4 and 8 mg l<sup>-1</sup>) were studied on callus growth and its secondary metabolites production. Results elucidated that after elicitation by 8 mg l<sup>-1</sup> of Ag NPs, significantly the highest callus growth rate (CGR) (0.02 mm day<sup>-1</sup>), callus fresh mass (CFM) (0.99 g), and carvacrol (0.68 mg l<sup>-1</sup>) and thymol (11.09 mg l<sup>-1</sup>) content was achieved. Comparing different *Thymus* species, notably the greatest carvacrol and thymol amount was obtained in *.kotschyanus* Boiss. & Hohen. and *T. Daenesis* Čelak. at 8 mg l<sup>-1</sup> concentration of Ag NPs. Hence, it is evident that the stimulation by NPs is dose-dependent. This study has potential to be commercially applied for the enhancement of pharmaceutical compounds in different species of *Thymus*.

**Key words:** Ag nanoparticles; *Thymus* species; *Zataria multiflora*; callus; carvacrol thymol

**Abbreviations:** 2,4-D: 2, 4-dichlorophenoxyacetic acid; Kin, kinetin; PGRs, plant growth regulators; HPLC, high performance liquid chromatography; NPs, nanoparticles

**Vpliv nanodelcev srebra (Ag) na fiziološke in biokemične lastnosti kalusa dveh vrst materine dušice (*Thymus* sp.) in vrste *Zataria multiflora* Boiss.**

**Izvleček:** Vrste iz rodu materine dušice (*Thymus* sp.) imajo velik pomen v proizvodnji hrane in zdravil. V raziskavi je bil preučevan potencialni učinek nanodelcev srebra kot eliciatorja na rast kalusa, tvorbo karvakrola in timola pri vrsti *Zataria multiflora* in treh vrstah materine dušice. Najprej je bil na Murashige in Skoog (MS) gojišču, ki je vsebovalo 2 mg l<sup>-1</sup> 2,4-D in 1 mg l<sup>-1</sup> kinetina, vzgojen kalus. Potem je bil preučevan učinek dveh različnih koncentracij srebrovih nanodelcev (4 in 8 mg l<sup>-1</sup>) na rast kalusa in in tvorbo sekundarnih metabolitov. Rezultati, pridobljeni z visokotlačno tekočinsko kromatografijo (HPLC) so pokazali, da je bila po uporabi 8 mg l<sup>-1</sup> srebrovih nanodelcev kot eliciatorjev dosežena značilno največja rast kalusa (CGR) (0,02 mm dan<sup>-1</sup>), največja sveža masa kalusa (CM) (0,99 g) in največja vsebnost karvakrola (0,68 mg l<sup>-1</sup>) in timola (11,09 mg l<sup>-1</sup>). V primerjavi različnih vrst materine dušice je bila dosežena največja vsebnost karvakrola in timola pri vrstah *T. kotschyanus* Boiss. & Hohen in *T. daenesis* Čelak pri koncentraciji srebrovih nanodelcev 8 mg l<sup>-1</sup>. Očitno je, da je stimulacijski učinek nanodelcev odvisen od doze. Izsledke raziskave bi lahko komercialno uporabili za povečanje tvorbe zdravilnih spojin pri različnih vrstah materine dušice.

**Ključne besede:** Ag nanodelci; vrste iz rodu *Thymus*; *Zataria multiflora*; kalus; karvakrol; timol

**Okrajšave:** 2,4-D: 2, 4-diklorfenoksi očetna kislina; Kin: kinetin; PGRs: rastlinski rastni regulatorji; HPLC: visokotlačna tekočinska kromatografija; NPs: nanodelci

1 Department of Agricultural Biotechnology, Payame Noor University (PNU), Tehran, Iran

2 Institute of Biotechnology, School of Agriculture, Shiraz University, Shiraz, Iran

3 Corresponding author, e-mail: nima.mosavat@gmail.com

4 Department of Engineering and Technology, Payame Noor University (PNU), Tehran, Iran

5 Department of Natural Resources, Isfahan University of Technology, Isfahan, Iran

6 Research Institute for Biotechnology and Bioengineering, Isfahan University of Technology, Isfahan, Iran

7 Department of Biotechnology, Quaid-i-Azam University, Islamabad, Pakistan

## 1 INTRODUCTION

Nanobiotechnology has enormous applications in diverse fields including agriculture, cosmetics, pharmaceuticals, and food industry (Kim et al., 2017; Rastogi et al., 2017). Nanoparticles (NPs) are employed as elicitors of various cell signaling pathways in metabolism of plants (Kim et al., 2017; Marslin, et al., 2017). NPs have been found to play crucial role in enhancement of plant secondary metabolites (SMs) by imposing oxidative stress and increasing cell membrane permeability (Jasim et al., 2017; Marslin et al., 2017; Ahmad et al., 2020). The effects of NPs on growth rate of plants (Sharma et al., 2012), germination of seeds (Zaka et al., 2016), production of SMs (Marslin et al., 2017; Mosavat et al., 2019; Zaka et al., 2016; Golkar et al., 2021) and plant physiology (Jasim et al., 2017; Sharma et al., 2012) have been studied recently. Although there are few studies related to effect of NPs on callus culture development, physiology and secondary metabolism (Dykman & Shchyogolev, 2017; Kokina et al., 2013; Marslin et al., 2017; Sanzari et al., 2019; Zuberza-Mena et al., 2017), still this research domain needs to be explored further. Silver (Ag) NPs possess unique properties in terms of toxicity and alteration of yield, development, antioxidant activities, and SMs production of plants due to their high catalysis and reactivity (Rastogi et al., 2017; Sadak 2019). Furthermore, the influence of Ag NPs on the callus cultures of *Solanum nigrum* L. (Ewais et al., 2015), *Prunella vulgaris* L. (Fazal et al., 2019) and *Caralluma tuberculata* N.E. Brown (Ali et al., 2019) has recently been studied.

*Thymus* L., belonging to Lamiaceae family, has world-wide distribution (Sajed et al., 2013). However, it is dominantly present in Asia, Europe and North Africa (Zarshenas & Krenn, 2015). The essential oils and SMs enhance the commercial value of flowers and leaves of *Thymus* making it a valuable crop in cosmetics, pharmaceuticals, and food and agriculture industry (Miraj & Kiani, 2016). A thyme-like plant, *Zataria multiflora* Boiss., belonging to Lamiaceae family, is wild plant found in only southern and central Pakistan, Afghanistan, and Iran (Sajed et al., 2013). Tissue culture propagation of thyme plant is well-known because it is a wide source of ingredients of pharmacology. Both *Thymus* sp. and *Z. multiflora* possess anti-cancerous, anti-inflammatory, anti-oxidant, anti-bacterial, anti-fungal, and anti-spasmodic properties (Mathela et al., 2010; Sajed et al., 2013). Naturally occurring terpenoid thymol (2-isopropyl-5-methyl phenol) and its phenol isomer, carvacrol/cymophenol are the phenolic compounds that play an important role in inducing these properties to *Thymus* sp. and *Z. multiflora* (Kianersi et al., 2021; Mathela et al., 2010). Some other properties like their use as additive in perfumes, deodor-

ant, toothpastes, soaps, etc. and as important flavoring agent in foods are also attributed to these compounds (Sajed et al., 2013).

The defense system of plants is activated by chemical, biological or physical elicitors (Asadollahei et al., 2022; Zhao et al., 2005). The gene expression is then modulated that transcribes the formation of SMs (Ajungla et al., 2009). A more efficient method could be the use of callus cultures for extracting SMs naturally from *Thymus* sp. and *Z. multiflora* in a sustained manner (Ramakrishna & Ravishankar, 2011). Previously, there is no report concerning the carvacrol and thymol production by imposing Ag NPs in cell cultures of *Thymus* sp. and *Z. multiflora*. Hence, the current study investigates proliferation of callus and SMs production in three *Thymus* species, i.e., *T. vulgaris* L., *T. daenensis* Čelak, and *T. kotschyanus* Boiss. & Hohen as well as *Z. multiflora* Boiss. after Ag NPs exposure.

## 2 MATERIALS AND METHODS

The seeds of *Thymus* species, i.e., *T. vulgaris*, *T. daenensis*, *T. Kotschyanus* and *Zataria multiflora* (two accessions) (Table 1) were deposited at Botanic Herbarium of Research Institute for Biotechnology and Bioengineering, Isfahan University of Technology (IUT), Isfahan, Iran, after collection from different geographical regions and identification by using Flora Iranica (Rechinger, 1982). Their characteristics is shown in Table 1. The seeds of four different species were surface sterilized with 70 % (v/v) ethanol for 1 min, followed by the addition of 3 % (v/v) sodium hypochlorite for 20 min, and then rinsing in sterile distilled water thrice. After surface sterilization, the seeds were grown in Murashige and Skoog (MS) (1962) medium (Duchefa, Netherland). These were incubated for germination and growth of plantlets. The leaflet explants from about 1-month old plantlets were cultured in MS medium containing 2,4-D (2 mg l<sup>-1</sup>) and Kin (1 mg l<sup>-1</sup>) supplemented with 3 % (w/v) sucrose (Sigma-Aldrich, USA), 0.8 % (w/v) agar (Sigma-Aldrich, USA) and 0.1 mg l<sup>-1</sup> myoinositol for callus induction. The pH was adjusted at 5.7. The samples were exposed to 16h/8h (light/dark) photoperiod for a period of 2 months at 23 ± 2 °C.

Silver (Ag) nano-powder was purchased from US Research Nanomaterials Inc., Houston, TX, USA having an average size of 30–50 nm and purity of 99.99 %. The nanoparticles were characterized by x-ray diffraction (XRD) and scanning electron microscopy (SEM) techniques by following the protocols of Javed et al. (2016). XRD was performed using Carlo ERBA Model EA 1108 analyzer and the instrument for getting SEM image was

**Table 1:** The geographical origins of *Thymus* sp. and *Z. multiflora* with their geographical traits collected from Iran

Species	Abbreviation	Origin	Genotype code	Latitude (m)	Longitude (m)	Altitude (m)
<i>Zataria multiflora</i> (1)	Zm (1)	Dehbala, Yazd, Iran	RIBB/ZM01/2016	31°59' N	54°11' E	2600
<i>Zataria multiflora</i> (2)	Zm (2)	Abadeh, Fars, Iran	RIBB/ZM02/2016	31°45' N	51°21' E	2030
<i>Thymus vulgaris</i>	Tv	Marvdasht, Fars, Iran	RIBB/TV01/2016	35°56' N	52°10' E	1620
<i>Thymus daenensis</i>	Td	Aligoodarz, Lorestan, Iran	RIBB/TD01/2016	33°24' E	49°41' E	2022
<i>Thymus kotschyanus</i>	Tk	Lahijan, Gilan, Iran	RIBB/TK01/2016	37°12' N	50°14' E	396

Hitachi S4800 (Japan). These NPs were added to MS medium after filter sterilization. The 2-months old friable callus (0.25 g) was transferred to MS containing 2,4-D ( $2 \text{ mg l}^{-1}$ ) and Kin ( $1 \text{ mg l}^{-1}$ ) under Ag NPs stress of 4 and  $8 \text{ mg l}^{-1}$ . This callus material was placed at  $23 \pm 2 \text{ }^\circ\text{C}$  under a photoperiod of 16h/8h. After incubation for 21 days of callus with Ag NPs, the callus growth rate (CGR) and callus fresh mass (CFM) was calculated. CGR was measured according to Afshar et al. (2016) every 7 days in 21 days period.

Later on, the quantity of carvacrol and thymol was obtained by high performance liquid chromatography (HPLC). The method of Castro et al. (2016) was utilized for preparing callus extracts. The process involved drying of 200 mg of callus from each treatment in an oven at  $50 \text{ }^\circ\text{C}$  for 24 h, and then soaking it in 5 ml of diethyl ether for a period of 24 h. In order to prevent the evaporation of diethyl ether, the vials were kept closed and extraction was performed in a cold room. After adding the 80 % of methanol (1 ml) to remaining solid material, the extracts were filtered ( $0.22 \text{ }\mu\text{m}$  pore size) into clean vials and prepared for injection to HPLC instrument. The HPLC (SY-8100 series, Beijing Beifan-Ruili Analytical Instrument, China) was performed by UV-VIS detector, a flow rate of  $0.9 \text{ ml min}^{-1}$ , injection volume of  $20 \text{ }\mu\text{l}$  at  $28 \text{ }^\circ\text{C}$ , C18 column ( $25 \text{ cm} \times 4.6 \text{ mm}$ , partial size  $5 \text{ }\mu\text{m}$ ), mobile phase methanol-water (80:20; v/v), and flow rate of  $0.9 \text{ ml min}^{-1}$ . The detection was carried out at 280 nm of wavelength and a pressure of 12 atm. The UV spectra of phenolic compounds were recorded at 280 and 320 nm. The content of carvacrol and thymol were determined based on the calibration curve of standard compounds, including carvacrol (Sigma-Aldrich, USA) and thymol (Sigma-Aldrich, USA). For this purpose, 4 concentrations (10, 25, 50, and  $100 \text{ mg l}^{-1}$ ) of carvacrol and 3 concentrations (25, 100, and  $400 \text{ mg l}^{-1}$ ) of thymol were examined by HPLC. The retention time for carvacrol and thymol were appeared at 3-4 min and 14-15 min, respectively. After calibration of the standards with HPLC, the quantities of carvacrol and thymol in different samples were calculated.

## 2.1 STATISTICAL ANALYSIS

The experimentation was conducted with three replications in completely randomized design and the statistics was determined using two-way analysis of variance (ANOVA). LSD test ( $p \leq 0.05$ ) in SAS software (SAS 9.1 Inc. USA) was applied to determine significant difference among the treatments.

## 3 RESULTS AND DISCUSSION

Callus growth and development as well as formation of secondary metabolic compounds is positively or negatively influenced by supplementing the growth medium with abiotic or biotic stress elicitors (Ajungla et al., 2009; Zaka et al., 2016). The contents of SMs show significant changes under elicitation of callus by different stresses (Fazal et al., 2016; Mosavat et al., 2019; Sanzari et al., 2019). NPs, specifically the metallic oxide NPs like ZnO, CuO,  $\text{TiO}_2$  act as oxidative abiotic stress elicitors (Lala 2021). According to Al-jibouri et al. (2012), thymol amount was increased by proline in *Origanum vulgare* L. Similarly, the production of significantly the highest content of hyperforin in *Hypericum perforatum* L. (Sharafi et al., 2013), rebaudioside A and stevioside in *Stevia rebaudiana* Bertoni (Javed, et al., 2018), and proline in *Triticum aestivum* L. (Barbasz, et al., 2016) under ZnO NPs elicitation has been previously documented.

Ag NPs show significant effect on the production of SMs in callus culture, resulting in their ultimate increase (Fazal et al., 2016). For instance, Ali et al. (2019) reported that various concentrations of Ag NPs significantly affected the callus proliferation and substantially increased the callus biomass and SMs in *Caralluma tuberculata*. In another study, Fazal et al. (2019) reported the positive effects of Ag NPs and Au NPs on the production of biomass and SMs in the cell culture of *Prunella vulgaris* L.

XRD of Ag NPs is given in Figure 1 which shows 100 % phase purity by the sharpness of peaks. Similar XRD pattern was obtained by Kim et al. (2006). The

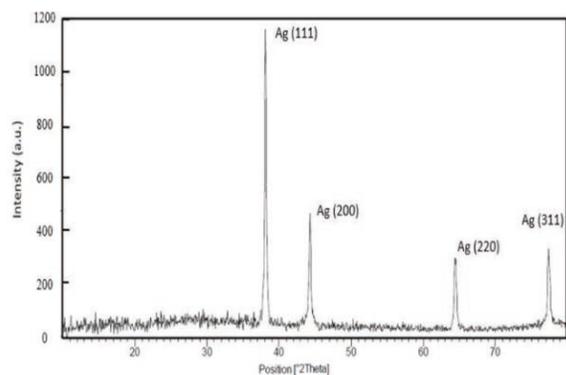


Figure 1: X-ray diffractogram (XRD) of Ag nanoparticles

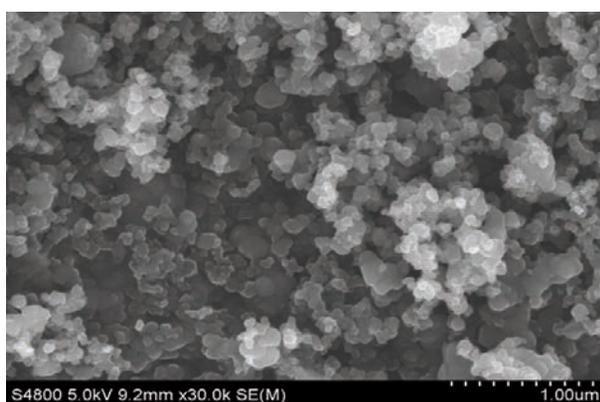


Figure 2: Scanning electron micrograph (SEM) of Ag nanoparticles

spherical shape of Ag NPs was illustrated by SEM image given in Figure 2 which is coinciding with the results of Elumalai et al. (2010).

The size, shape, surface, concentration and chemical composition of NPs cause stimulatory or inhibitory effects on the growth of callus cells (Al-Jibouri et al., 2012). The synthesis and accumulation of SMs in cells is enhanced by an increased surface area of NPs as a result of reduced size and transport of NPs in to the cells apo-

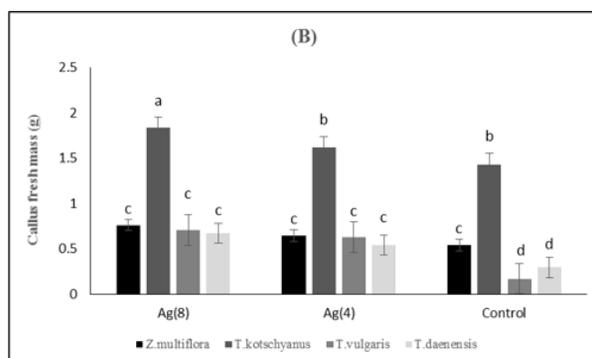
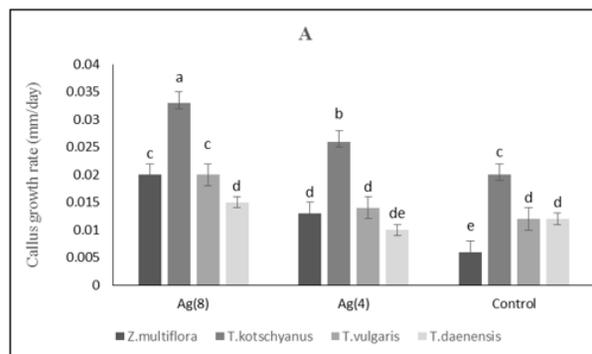


Fig 3: Effect of Ag nanoparticles on callus growth rate (A) and callus fresh mass (B) content of different *Thymus* species and *Z. multiflora* under callus culture

plastically which increases electrostatic interactions between living cell membranes (Javed et al., 2017). Table 2 and Figure 3 shows the effect of Ag NPs on callus growth of *Thymus* sp. and *Zataria multiflora*.

Compact calli with white and greenish colour were obtained after 10 days upon control culture (no AgNPs), whereas friable watery calli with white, greenish or yellowish colour were observed after 10-13 days upon culture supplemented with AgNPs (Figure 4.)

A significant effect is produced on callus traits by different concentrations of Ag NPs in this study that is coherent with the reports about effects of Ag NPs on

Table 2: Effect of different concentrations of Ag NPs on color, texture, growth rate, and fresh mass of callus cultures of *Thymus* sp. and *Zataria multiflora*

Nanoparticle	Concentration (mg l <sup>-1</sup> )	Color and Texture	CGR <sup>I</sup> (mm day <sup>-1</sup> )				CFM <sup>II</sup> (g)
			7	14	21	Mean	
Ag	8	Green, friable	0.045	0.06	0.015	0.022 <sup>a</sup>	0.99 <sup>a</sup>
Ag	4	Green, friable	0.07	0.06	0.032	0.015 <sup>b</sup>	0.85 <sup>ab</sup>
Control	-	White to yellow, soft	0.02	0.01	0.006	0.012 <sup>c</sup>	0.61 <sup>b</sup>

Mean values followed by the same letter in each column are not significantly different at  $p < 0.05$  (Least Significant Difference Test). I: CGR: Callus growth rate, II: CFM: Callus fresh mass

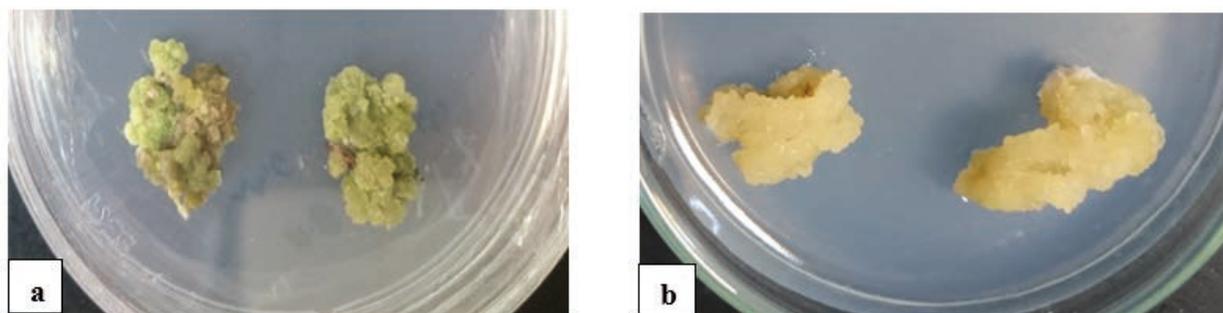


Fig 4: Friable calli with greenish or yellowish colour of *Thymus* species and *Z. multiflora* after culture treatment with AgNP<sub>5</sub> (a) and under control culture (b)

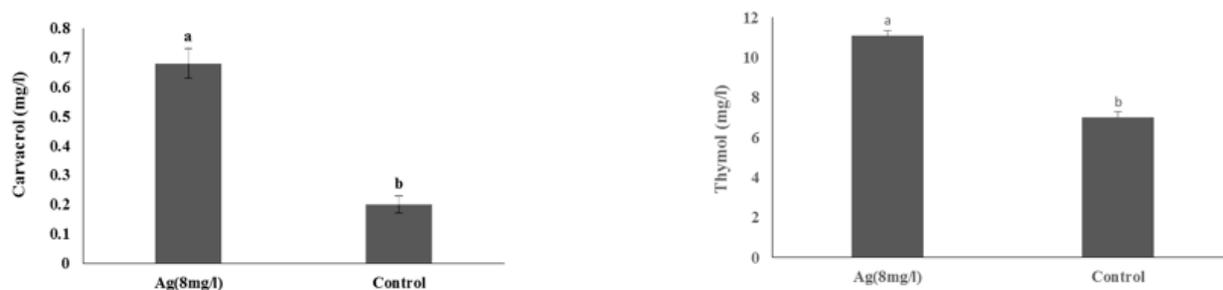


Fig 5: Effect of 8 mg l<sup>-1</sup> Ag NPs on *in vitro* production of carvacrol and thymol in *Thymus* sp. and *Zataria multiflora*

callus culture of *Solanum nigrum* L. (Ewais et al., 2015) which gain friable watery calli with greenish or yellowish colour were observed after 10-13 days upon culture supplemented with AgNPs and effects of TiO<sub>2</sub> NPs on the callus of *Hordeum vulgare* L. (Mandeh et al., 2012). The higher TiO<sub>2</sub> NPs concentration influenced callogenesis of *Hordeum vulgare* explants in this study. Also according to Kokina et al. (2013) elicitation with Ag and Au NPs shows positive effects on callus width and length in *Linum usitatissimum* L. The positive effects of different NPs on callus growth of *Prunella vulgaris* L. is reported by Fazal et al. (2019). Callus growth traits affected by ZnO NPs in *Solanum lycopersicum* Mill. have also been reported (Alharby et al., 2016).

The effects of elicitation by Ag NPs on production of carvacrol and thymol under *in vitro* conditions are presented in Figure 5. The production of thymol and carvacrol was determined at 8 mg l<sup>-1</sup> concentration of Ag NPs and control treatment. The carvacrol (0.68 mg l<sup>-1</sup>) and thymol (11.09 mg l<sup>-1</sup>) quantity was enhanced under 8 mg l<sup>-1</sup> of Ag NPs. The chromatographic separation of the methanolic extracts in *Zataria multiflora* for carvacrol and thymol by HPLC is given (Figure 6). Asadollahei et al. (2022) employed different concentrations of CuNPs in *in vitro* culture medium and observed significant rise in thymol and carvacrol content compared to control in *Zataria multiflora*. This study elucidated that the selection

of appropriate plant species and suitable elicitor is crucial for increasing the production of bioactive compounds as well as antioxidants of *Zataria multiflora*. This can be done by inducing expression changes in the biosynthetic pathways of thymol and carvacrol. In fact, the gene expression patterns of the pathways of formation of thymol and carvacrol were greatly influenced by the Ag NPs in our study which is the phenomenon well explained by the studies of Kianersi et al. (2021).

The interactive effects of NPs and genotypes/species of *Thymus* and *Zataria multiflora* for production of SMs

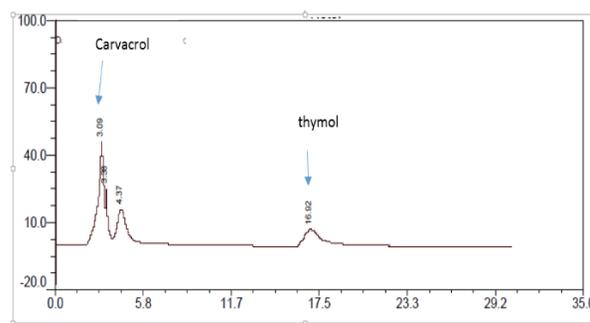


Fig 6: Representative HPLC chromatograms of thymol and carvacrol of *Z. multiflora*. Peak identifications were performed by matching retention time and UV spectra against commercially available reference compounds

have been presented in Table 3. The exposure of Ag NPs has significantly increased the content of two SMs of callus compared to control. Moreover, *in vitro* synthesis of thymol was notably greater than that of carvacrol which is also evident from the molecular studies of Kianersi et al. (2021). The highest content of carvacrol ( $1.06 \text{ mg l}^{-1}$ ) was observed at  $8 \text{ mg l}^{-1}$  concentration of Ag NPs in *T. kotschyanus*, whereas the least amount ( $0.10 \text{ mg l}^{-1}$ ) was observed in control treatment of *T. daenensis*. Furthermore, the highest concentration of thymol was obtained at  $8 \text{ mg l}^{-1}$  of Ag NPs in callus of *T. daenensis* ( $19.75 \text{ mg l}^{-1}$ ), while the least thymol content ( $3.95 \text{ mg l}^{-1}$ ) was achieved in *T. daenensis* under control condition.

This result can be well supported by the phenomenon that NPs trigger thymol synthetic pathways and/or transcription factors more than the carvacrol pathways (Mosavat et al., 2019). Taking into account of concurrent studies, the phenolics and flavonoids production is activated by ZnO NPs in seedlings of *Brassica nigra* L. (Zafar et al., 2016) Additionally, the significant rise in hyperforin content in cell suspension culture of *Hypericum perforatum* L. under ZnO NPs stress is reported (Sharafi et al., 2013). A complex variety of elicitation effects on *in vitro* synthesis of SMs is obtained using different types of elicitors (Goswami et al., 2017; Marslin et al., 2017; Syu et al., 2014), plant tissues (Ajungla et al., 2009), and physicochemical environment of various species (Shakya et al., 2019).

#### 4 CONCLUSION

The formation of callus from *Thymus* species and *Zataria multiflora* was performed in the presence of Ag NPs elicitors. Addition of abiotic elicitors, i.e., Ag NPs ( $8$

**Table 3:** The effect of species  $\times$  NPs interaction on *in vitro* production of thymol and carvacrol in callus culture of *Thymus* species and *Z. multiflora*

Species	Nanoparticles ( $\text{mg l}^{-1}$ )	Thymol ( $\text{mg l}^{-1}$ )	Carvacrol ( $\text{mg l}^{-1}$ )
<i>Z. multiflora</i>	Ag (8)	$12.76^b \pm 0.02$	$1.05^a \pm 0.03$
	Control	$7.06^g \pm 0.01$	$0.20^{bc} \pm 0.01$
<i>T. kotschyanus</i>	Ag (8)	$7.90^e \pm 0.01$	$1.06^a \pm 0.04$
	Control	$7.39^f \pm 0.01$	$0.26^b \pm 0.01$
<i>T. vulgaris</i>	Ag (8)	$11.06^c \pm 0.04$	$0.29^b \pm 0.01$
	Control	$9.64^d \pm 0.04$	$0.10^c \pm 0.002$
<i>T. daenensis</i>	Ag (8)	$19.75^a \pm 0.02$	$0.31^b \pm 0.01$
	Control	$3.95^h \pm 0.03$	$0.23^{bc} \pm 0.02$

Each value represents Mean  $\pm$  SE. Mean values followed by the same letter are not significantly different at  $p < 0.05$  (Least Significant Difference Test)

$\text{mg l}^{-1}$ ) to the MS medium played a vital role in enhancing the thymol and carvacrol content in the callus cultures of different *Thymus* species and *Zataria multiflora*. In other words, Ag nano-elicitors applied to the *in vitro* callus cultures of *Thymus* species and *Zataria multiflora* in our study resulted in increase in SMs production at a concentration of  $8 \text{ mg l}^{-1}$ . Our finding opens the way for studies involving relationship between chemical elicitors and formation & accumulation of thymol/carvacrol. In future, transcriptomic and metabolomics studies should be performed to elucidate the regulation of SMs production under an elicitation of Ag NPs in these medicinal plants.

#### 5 ACKNOWLEDGMENT

The authors wish to express their gratitude to the Research Institute for Biotechnology and Bioengineering, Isfahan University of Technology, Isfahan, Iran and Payame Noor University, Isfahan, Iran.

#### 6 DECLARATION OF CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

#### 7 REFERENCES

- Afshar, B., & Golkar, P. (2016). Mucilage synthesis by *in vitro* cell culture in different species of *Alyssum*. *BioTechnologia. Journal of Biotechnology Computational Biology and Bionanotechnology*, 97(2), 79–86. <https://doi.org/10.5114/bta.2016.60778>
- Ahmad, M. A., Javed, R., Adeel, M., Rizwan, M., Ao, Q., & Yang, Y. (2020). Engineered ZnO and CuO nanoparticles ameliorate morphological and biochemical response in tissue culture regenerants of candyleaf (*Stevia rebaudiana*). *Molecules*, 25(6), 1356. <https://doi.org/10.3390/molecules25061356>
- Ajungla, L., Patil, P. P., Barmukh, R. B., & Nikam, T. D. (2009). Influence of biotic and abiotic elicitors on accumulation of hyoscyamine and scopolamine in root cultures of *Datura metel* L. *Indian Journal of Biotechnology*, 8(3), 317–322.
- Alharby, H. F., Metwali, E. M. R., Fuller, M. P., & Aldhebani, A. Y. (2016). Impact of application of zinc oxide nanoparticles on callus induction, plant regeneration, element content and antioxidant enzyme activity in tomato (*Solanum lycopersicum* mill.) under salt stress. *Archives of Biological Sciences*, 68(4), 723–735. <https://doi.org/10.2298/ABS151105017A>
- Ali, A., Mohammad, S., Khan, M. A., Raja, N. I., Arif, M., Kamil,

- A., & Mashwani, Z. ur R. (2019). Silver nanoparticles elicited *in vitro* callus cultures for accumulation of biomass and secondary metabolites in *Caralluma tuberculata*. *Artificial Cells, Nanomedicine and Biotechnology*, 47(1), 715–724. <https://doi.org/10.1080/21691401.2019.1577884>
- Al-Jibouri, A. M. J., Abd, A. S., Majeed, D. M., & Ismail, E. N. (2012). Influence of abiotic elicitors on accumulation of thymol in callus cultures of *Origanum vulgare* L. *Journal of Life Sciences*, 6(10), 1094.
- Asadollahei, M. V., Yousefifard, M., Tabatabaeian, J., Nekonam, M. S., & Mahdavi, S. M. E. (2022). Effect of elicitors on secondary metabolites biosynthesis in *Zataria multiflora* Boiss. *Industrial Crops and Products*, 181, 114789. <https://doi.org/10.1016/j.indcrop.2022.114789>
- Barbasz, A., Kreczmer, B., & Oćwieja, M. (2016). Effects of exposure of callus cells of two wheat varieties to silver nanoparticles and silver salt (AgNO<sub>3</sub>). *Acta Physiologiae Plantarum*, 38(3), 1–11. <https://doi.org/10.1007/s11738-016-2092-z>
- Castro, A. H. F., Braga, K. de Q., de Sousa, F. M., Coimbra, M. C., & Chagas, R. C. R. (2016). Callus induction and bioactive phenolic compounds production from *Byrsonima verbascifolia* (L.) DC. (Malpighiaceae). *Revista Ciencia Agronomica*, 47(1), 143–151. <https://doi.org/10.5935/1806-6690.20160017>
- Dykman, L. A., & Shchyogolev, S. Y. (2017). Interactions of plants with noble metal nanoparticles. *Agricultural Biology*, 52(1), 13. <https://doi.org/10.15389/agrobiol.2017.1.13eng>
- Elumalai, E. K., Prasad, T. N. V. K. V., Hemachandran, J., Vivian Therasa, S., Thirumalai, T., & David, E. (2010). Extracellular synthesis of silver nanoparticles using leaves of *Euphorbia hirta* and their antibacterial activities. *Journal of Pharmaceutical Sciences and Research*, 2(9), 549–554.
- Ewais, E. ., Desouky, S., & Elshazly, E. . (2015). Evaluation of callus responses of *Solanum nigrum* L. exposed to biologically synthesized silver nanoparticles. *Nanoscience and Nanotechnology*, 5(3), 45–56.
- Fazal, H., Abbasi, B. H., Ahmad, N., & Ali, M. (2016). Elicitation of medicinally important antioxidant secondary metabolites with silver and gold nanoparticles in callus cultures of *Prunella vulgaris* L. *Applied Biochemistry and Biotechnology*, 180(6), 1076–1092. <https://doi.org/10.1007/s12010-016-2153-1>
- Fazal, H., Abbasi, B. H., Ahmad, N., Ali, M., Shujait Ali, S., Khan, A., & Wei, D. Q. (2019). Sustainable production of biomass and industrially important secondary metabolites in cell cultures of selfheal (*Prunella vulgaris* L.) elicited by silver and gold nanoparticles. *Artificial Cells, Nanomedicine and Biotechnology*, 47(1), 2553–2561. <https://doi.org/10.1080/21691401.2019.1625913>
- Golkar, P., Bakhtiari, M.A. & Bazarganipour, M., (2021). The effects of nanographene oxide on the morpho-biochemical traits and antioxidant activity of *Lepidium sativum* L. under *in vitro* salinity stress. *Scientia Horticulturae*, 288, p.110301. <https://doi.org/10.1016/j.scienta.2021.110301>
- Goswami, L., Kim, K. H., Deep, A., Das, P., Bhattacharya, S. S., Kumar, S., & Adelodun, A. A. (2017). Engineered nanoparticles: Nature, behavior, and effect on the environment. *Journal of Environmental Management*, 196, 297–315. <https://doi.org/10.1016/j.jenvman.2017.01.011>
- Jasim, B., Thomas, R., Mathew, J., & Radhakrishnan, E. K. (2017). Plant growth and diosgenin enhancement effect of silver nanoparticles in Fenugreek (*Trigonella foenum-graecum* L.). *Saudi Pharmaceutical Journal*, 25(3), 443–447. <https://doi.org/10.1016/j.jsps.2016.09.012>
- Javed, R., Ahmed, M., Haq, I. ul, Nisa, S., & Zia, M. (2017). PVP and PEG doped CuO nanoparticles are more biologically active: Antibacterial, antioxidant, antidiabetic and cytotoxic perspective. *Materials Science and Engineering C*, 79, 108–115. <https://doi.org/10.1016/j.msec.2017.05.006>
- Javed, R., Usman, M., Tabassum, S., & Zia, M. (2016). Effect of capping agents: Structural, optical and biological properties of ZnO nanoparticles. *Applied Surface Science*, 386, 319–326. <https://doi.org/10.1016/j.apsusc.2016.06.042>
- Javed, R., Yucesan, B., Zia, M., & Gurel, E. (2018). Elicitation of secondary metabolites in callus cultures of *Stevia rebaudiana* Bertoni grown under ZnO and CuO nanoparticles stress. *Sugar Tech*, 20(2), 194–201. <https://doi.org/10.1007/s12355-017-0539-1>
- Kianersi, F., Pour-Aboughadareh, A., Majdi, M., & Poczai, P. (2021). Effect of methyl jasmonate on thymol, carvacrol, phytochemical accumulation, and expression of key genes involved in thymol/carvacrol biosynthetic pathway in some Iranian Thyme Species. *International Journal of Molecular Sciences*, 22(20), 11124. <https://doi.org/10.3390/ijms222011124>
- Kim, D. H., Gopal, J., & Sivanesan, I. (2017). Nanomaterials in plant tissue culture: the disclosed and undisclosed. *RSC advances*, 7(58), 36492–36505. <https://doi.org/10.1039/C7RA07025J>
- Kim, D., Jeong, S., & Moon, J. (2006). Synthesis of silver nanoparticles using the polyol process and the influence of precursor injection. *Nanotechnology*, 17(16), 4019–4024. <https://doi.org/10.1088/0957-4484/17/16/004>
- Kokina, I., Gerbreders, V., Sledevskis, E., & Bulanovs, A. (2013). Penetration of nanoparticles in flax (*Linum usitatissimum* L.) calli and regenerants. *Journal of Biotechnology*, 165(2), 127–132. <https://doi.org/10.1016/j.jbiotec.2013.03.011>
- Lala, S. (2021). Nanoparticles as elicitors and harvesters of economically important secondary metabolites in higher plants: A review. *IET Nanobiotechnology*, 15(1), 28–57. <https://doi.org/10.1049/nbt2.12005>
- Mandeh, M., Omidi, M., & Rahaie, M. (2012). In Vitro influences of TiO<sub>2</sub> nanoparticles on barley (*Hordeum vulgare* L.) tissue culture. *Biological Trace Element Research*, 150(1–3), 376–380. <https://doi.org/10.1007/s12011-012-9480-z>
- Marslin, G., Sheeba, C. J., & Franklin, G. (2017). Nanoparticles alter secondary metabolism in plants via ROS burst. *Frontiers in Plant Science*, 8, 832. <https://doi.org/10.3389/fpls.2017.00832>
- Mathela, C. S., Singh, K. K., & Gupta, V. K. (2010). Synthesis and *in vitro* antibacterial activity of thymol and carvacrol derivatives. *Acta Poloniae Pharmaceutica - Drug Research*, 67(4), 375–380.
- Miraj, S., & Kiani, S. (2016). Study of pharmacological effect of *Ocimum basilicum*: A review. *Der Pharmacia Lettre*, 8(9), 315–320.

- Mosavat, N., Golkar, P., Yousefifard, M., & Javed, R. (2019). Modulation of callus growth and secondary metabolites in different *Thymus* species and *Zataria multiflora* micro-propagated under ZnO nanoparticles stress. *Biotechnology and Applied Biochemistry*, 66(3), 316–322. <https://doi.org/10.1002/bab.1727>
- Murashige, T., & Skoog, F. (1962). A Revised Medium for Rapid Growth and Bio Assays with Tohaoco Tissue Cultures. *Physiologia Plantarum*, 15(3), 474–497. <https://doi.org/10.1111/j.1399-3054.1962.tb08052.x>
- Ramakrishna, A., & Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signaling and Behavior*, 6(11), 1720–1731. <https://doi.org/10.4161/psb.6.11.17613>
- Rastogi, A., Zivcak, M., Sytar, O., Kalaji, H. M., He, X., Mbarkki, S., & Brestic, M. (2017). Impact of metal and metal oxide nanoparticles on plant: A critical review. *Frontiers in Chemistry*, 5, 78. <https://doi.org/10.3389/fchem.2017.00078>
- Rechinger, K. (1982). Flora Iranica, Vol. 150. *IC Hedge - Graz: Akademische Druck Und Verlagsanstalt*, 150, 543–544.
- Sadak, M. S. (2019). Impact of silver nanoparticles on plant growth, some biochemical aspects, and yield of fenugreek plant (*Trigonella foenum-graecum*). *Bulletin of the National Research Centre*, 43(1), 1-6. <https://doi.org/10.1186/s42269-019-0077-y>
- Sajed, H., Sahebkar, A., & Iranshahi, M. (2013). *Zataria multiflora* Boiss. (Shirazi thyme) - An ancient condiment with modern pharmaceutical uses. *Journal of Ethnopharmacology*, 145(3), 686–698. <https://doi.org/10.1016/j.jep.2012.12.018>
- Sanzari, I., Leone, A., & Ambrosone, A. (2019). Nanotechnology in plant science: to make a long story short. *Frontiers in Bioengineering and Biotechnology*, 7, 120. <https://doi.org/10.3389/fbioe.2019.00120>
- Shakya, P., Marslin, G., Siram, K., Beerhues, L., & Franklin, G. (2019). Elicitation as a tool to improve the profiles of high-value secondary metabolites and pharmacological properties of *Hypericum perforatum*. *Journal of Pharmacy and Pharmacology*, 71(1), 70–82. <https://doi.org/10.1111/jphp.12743>
- Sharafi, E., Fotokian, M. H., & Loo, H. (2013). Improvement of hypericin and hyperforin production using zinc and iron nano-oxides as elicitors in cell suspension culture of John'swort (*Hypericum perforatum* L.). *Journal of Medicinal Plants and By-products*, 2(2).
- Sharma, P., Bhatt, D., Zaidi, M. G. H., Saradhi, P. P., Khanna, P. K., & Arora, S. (2012). Silver nanoparticle-mediated enhancement in growth and antioxidant status of *Brassica juncea*. *Applied Biochemistry and Biotechnology*, 167(8), 2225–2233. <https://doi.org/10.1007/s12010-012-9759-8>
- Syu, Y. yu, Hung, J. H., Chen, J. C., & Chuang, H. wen. (2014). Impacts of size and shape of silver nanoparticles on Arabidopsis plant growth and gene expression. *Plant Physiology and Biochemistry*, 83, 57–64. <https://doi.org/10.1016/j.plaphy.2014.07.010>
- Zafar, H., Ali, A., Ali, J. S., Haq, I. U., & Zia, M. (2016). Effect of ZnO nanoparticles on *Brassica nigra* seedlings and stem explants: Growth dynamics and antioxidative response. *Frontiers in Plant Science*, 7, 535. <https://doi.org/10.3389/fpls.2016.00535>
- Zaka, M., Abbasi, B. H., Rahman, L.-U., Shah, A., & Zia, M. (2016). Synthesis and characterisation of metal nanoparticles and their effects on seed germination and seedling growth in commercially important *Eruca sativa*. *IET Nanobiotechnology*, 10(3), 1–7. <https://doi.org/10.1049/iet-nbt.2015.0039>
- Zarshenas, M. M., & Krenn, L. (2015). A critical overview on *Thymus daenensis* Celak.: phytochemical and pharmacological investigations. *Journal of Integrative Medicine*, 13(2), 91–98. [https://doi.org/10.1016/S2095-4964\(15\)60166-2](https://doi.org/10.1016/S2095-4964(15)60166-2)
- Zhao, J., Davis, L., & Verpoorte, R. (2005). Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnology Advances*, 23(4), 283–333. <https://doi.org/10.1016/j.biotechadv.2005.01.003>
- Zuverza-Mena, N., Martínez-Fernández, D., Du, W., Hernandez-Viezcas, J. A., Bonilla-Bird, N., López-Moreno, M. L., Gardea-Torresdey, J. L. (2017). Exposure of engineered nanomaterials to plants: Insights into the physiological and biochemical responses-A review. *Plant Physiology and Biochemistry*, 110, 236–264. <https://doi.org/10.1016/j.plaphy.2016.05.037>