Sprejeto (accepted): 2003-10-30

# Growth and root respiration of C4 plants under CO, enrichment

Rast in dihanje korenin C4 rastlin pri povečani koncentraciji CO,

Irena MAČEK,1 Hardy PFANZ,2 Dominik VODNIK1

<sup>1</sup>University of Ljubljana, BF, Agronomy Dep., Jamnikarjeva 101, SI-1001 Ljubljana, Slovenia; E-mail: irena.macek@bf.uni-lj.si

<sup>2</sup>Institut für Angewandte Botanik, Universität Essen, D-45117 Essen, FRG

**Abstract.** Respiratory measurements of apical root parts of several C4 plant species (*Echinochloa crus-galli* var. *crus-galli*, *Setaria pumila* and *Zea mays* DK 312 (Dekalb, USA) subjected to an elevated CO<sub>2</sub> regime during growth in climatic chambers or at natural CO<sub>2</sub> springs were performed.

Biomass production, root respiratory potential and root respiration of *Echinochloa* was not significantly changed by high atmospheric CO<sub>2</sub> treatment in the climatic chambers, compared to ambient CO<sub>2</sub> treatment.

Root respiratory potential of C4 weeds (*Echinochloa crus-galli* and *Setaria pumila*) growing in natural  $CO_2$  spring area was not significantly affected by extremely high  $CO_2$  in the rhizosphere. Yet, respiratory potential of one and a half month old sown maize seedlings was significantly lower in the roots exposed to naturally elevated  $CO_2$  concentrations.

**Key words:** root respiration, ETS activity, respiratory potential, C4 plants, *Echinochloa crus-galli*, *Zea mays*, *Setaria pumila*, elevated CO<sub>2</sub>, natural CO<sub>2</sub> springs, CO<sub>2</sub> mofette

**Izvleček.** V pričujoči raziskavi smo merili dihalno aktivnost apikalnih delov korenin nekaterih C4 vrst (navadne kostrebe *Echinochloa crus-galli* var. *crus-galli*, sivozelenega muhviča *Setaria pumila* in koruze *Zea mays* DK 312 (Dekalb, ZDA), izpostavljenih povečani koncentraciji CO<sub>2</sub> med rastjo v klimatskih komorah ali ob naravnih izvirih CO<sub>2</sub>.

Pri navadni kostrebi povečana koncentracija CO<sub>2</sub> v klimatskih komorah ni značilno vplivala na produkcijo biomase, dihalni potencial in dihanje korenin.

Ekstremno povečana koncentracija CO<sub>2</sub> v rizosferi rastlin (navadne kostrebe in sivozelenega muhviča) ob naravnih izvirih CO<sub>2</sub> ni značilno vplivala na dihalni potencial v koreninah. Ta je bil značilno manjši le pri mesec in pol stari sejani koruzi rastoči na področju naravnih izvirov CO<sub>2</sub>.

Ključne besede: dihanje korenin, aktivnost ETS, dihalni potencial, C4 rastline, Echinochloa crus-galli, Zea mays, Setaria pumila, povečana koncentracija CO<sub>2</sub>, naravni izviri CO<sub>2</sub>, CO<sub>2</sub> mofeta

### Introduction

An elevated atmospheric  $\mathrm{CO}_2$  concentration can have a significant effect on growth and carbon metabolism of many plant species. On a daily basis, more than 50% of the photosynthates produced, may be simultaneously respired by roots (LAMBERS et al. 2002). Compared to the number of papers on inhibition of aboveground shoot respiration by elevated  $\mathrm{CO}_2$  (e.g. AMTHOR 1991, DRAKE et al. 1997, TJOELKER et al. 2001), very little information on the effects of elevated  $\mathrm{CO}_2$  on root respiration has been published so far. Furthermore, the reported  $\mathrm{CO}_2$  effects on root respiration are rather heterogeneous, limited to only few plant species and differently discussed among authors (e.g. BURTON et al. 1997, YODER et al. 2000, LAMBERS et al. 2002). LAMBERS et. al. (1996) report that there is insufficient evidence to state that root respiration is inhibited by the  $[\mathrm{CO}_2]$  around roots in a similar manner as leaf respiration is inhibited by elevated  $\mathrm{CO}_2$  in the atmosphere. Moreover, there is no convincing evidence for a direct effect of elevated atmospheric  $\mathrm{CO}_2$  on the specific rate of root respiration or the fraction of carbon required for root respiration. However, there are probable indirect effects of elevated  $\mathrm{CO}_2$  on the carbon requirement of plants in natural systems.

The partial pressure of  $\mathrm{CO}_2$  in soil may differ from the  $\mathrm{CO}_2$  partial pressure above ground and it seems to be more variable. The concentration of  $\mathrm{CO}_2$  in soil rapidly increases after rainfall because its diffusion through the gas phase is restricted by the water saturation of the soil. Thus, plant roots are frequently exposed to relatively high  $\mathrm{CO}_2$  concentrations. Water flooding can also present a longer exposure of roots to elevated  $\mathrm{CO}_2$  and consequently hypoxic environment. Plants growing near  $\mathrm{CO}_2$  springs are exposed to extreme  $\mathrm{CO}_2$  regimes that can also lead to hypoxic conditions in the rhizosphere. Vegetation at  $\mathrm{CO}_2$  springs has been exposed to such conditions for long periods, giving time for acclimation, and perhaps also genetic adaptation of plants (VODNIK et al. 2002). Thus, natural  $\mathrm{CO}_2$  springs may represent a good model ecosystem to study effects of elevated  $\mathrm{CO}_2$  on plants (RASCHI et al. 1997, BADIANI et al. 1999).

Our work was performed on three C4 plant species *Echinochloa crus-galli* var. *crus-galli*, *Setaria pumila* and *Zea mays* DK 312 (Dekalb, USA) subjected to an elevated CO<sub>2</sub> regime during growth in climatic chambers as well as in a CO<sub>2</sub> spring-area Stavešinci in NE Slovenia.

#### Material and methods

Seedlings of *Echinochloa*, the offsprings of plants growing near the  $CO_2$  springs, were grown in climatic chambers under ambient (368.4 ± 15.6  $\mu$ mol  $CO_2$  mol<sup>-1</sup>) and elevated (1906.2 ± 195.0  $\mu$ mol  $CO_2$  mol<sup>-1</sup>)  $CO_2$  for seven weeks at the temperature 25 °C during the light (14 h) and 16 °C during the dark period (10 h) of the day. The fumigation started two weeks after seed germination. After the growth period plant roots were sampled for respiratory measurements.

In the July 2002, root samples were also collected for natural growing *Echinochloa*, *Setaria* and one and a half month old plants of sown maize (*Zea mays* DK 312, Dekalb, USA) growing in the natural CO<sub>2</sub> spring area Stavešinci in NE Slovenia (for detailed information on the site characteristics and other details see VODNIK et al. in press). The plants were chosen according to their height and the preliminary measured soil CO<sub>2</sub> concentration in the rooting zone by a gas analyzer GA 2000 (Ansyco, FRG). A significant correlation between plant height and plant exposition to elevated CO<sub>2</sub> from CO<sub>2</sub> springs is known from our previous studies (VODNIK et al. 2002<sup>a,b</sup>, TURK et al. 2002).

Root respiration rates were measured as oxygen consumption on root tip segments (1 cm length) using Clark-type oxygen electrodes (Hansatech, Norfolk, UK). Measurements were performed in 50 mM MES (morpholinoethanesulfonic acid) buffer solution pH 6 at 20 °C.

The root respiratory potential - electron transport system (ETS) activity was determined on root tip segments (1 cm length) using the iodonitrotetrazolium salt (INT) method described by KENNER & AHMED (1975).

Statistical analyses were performed by Statgraphic Plus 4.0 (Statistical Graphics Corp.).

## Results and discussion

In the fumigation experiment no significant impact of high atmospheric CO<sub>2</sub> concentration on biomass production of *Echinochloa* was found (Tab. 1). In general, the growth response of C4 plants to elevated [CO<sub>2</sub>] is inconsistent and depends on various environmental factors (GHANNOUM et al. 2000). For *Echinochloa* the increase of plant biomass under elevated CO<sub>2</sub> concentration (690 µmol mol<sup>-1</sup>) was documented by ZISKA et al. (1997).

Measurements of root oxygen consumption revealed no significant differences in root respiration of elevated  ${\rm CO_2}$  chamber-grown *Echinochloa* plants. There was also no significant difference in root respiratory potential (Tab. 1).

Table 1: Biomass production, root respiration and root respiratory potential of *E. crus-galli* var. *crus-galli* subjected to elevated atmospheric CO<sub>2</sub> in climatic chambers, (avg ±SD).

|                             | Ambient CO <sub>2</sub>                                    | Elevated CO <sub>2</sub>                                 |
|-----------------------------|------------------------------------------------------------|----------------------------------------------------------|
|                             | $(368.4 \pm 15.6  \mu \text{mol CO}_{2}  \text{mol}^{-1})$ | (1906.2 ± 195.0 μmol CO <sub>2</sub> mol <sup>-1</sup> ) |
| Dry weight of shoot (g) (1) | $0.95 \pm 0.26$                                            | $1.15 \pm 0.26$                                          |
| Dry weight of root (g) (1)  | $0.83 \pm 0.16$                                            | $0.89 \pm 0.33$                                          |
| Root respiration(2)         | $1.5 \pm 0.6$                                              | $1.8 \pm 0.5$                                            |
| ETS roots(3)                | $1.0 \pm 0.2$                                              | $1.0 \pm 0.2$                                            |

<sup>&</sup>lt;sup>(1)</sup>By ANOVA, n = 24. <sup>(2)</sup>Given as nmol  $O_2$  g<sup>-1</sup> fresh wt s<sup>-1</sup>, by ANOVA, n = 15. <sup>(3)</sup>Given as  $\mu$ g  $O_2$  g<sup>-1</sup> fresh wt h<sup>-1</sup>, by ANOVA, n = 15

In addition to these findings, measurements of *Echinochloa* exposed to different CO<sub>2</sub> regimes at the natural CO<sub>2</sub> spring Stavešinci showed no significant effects of high rhizospheric CO<sub>2</sub> concentration on root respiratory potential of the root-tip segments. The same was true for another C4 plant *Setaria pumila* (Tab. 2). It is to conclude that both species are relatively insensitive to high CO<sub>2</sub>. A high tolerance of *E. crus-galli* to hypoxia is known from different studies and is especially well documented for its variety *E. crus-galli* var. *oryzicola* (BUCHANAN & al. 2000). Germination of *E. crus-galli* could be stimulated by elevated CO<sub>2</sub> as it was shown by YOSHIOKA & al. (1998). In this study germination was stimulated by exposure to 30 mmol mol<sup>-1</sup> CO<sub>2</sub> and it was concluded that soil CO<sub>2</sub> is responsible for causing intermittent flushes of seed germination of this species after heavy rainfall. This could also explain the presence of germinating and growing *Echinochloa* plants at the sites

with extreme CO<sub>2</sub> concentrations in the natural CO<sub>2</sub> spring Stavešinci as reported by KALIGARIČ (2001). No similar reports have been published on *Setaria pumila*.

Table 2: Shoot height and root respiratory potential of *E. crus-galli* var. *crus-galli*, *S. pumila* and *Z. mays* subjected to elevated soil and atmospheric CO<sub>2</sub> at a natural CO<sub>2</sub> spring (avg  $\pm$  SD).

| Plant species          | CO, exposure(1) | Mean height (cm) <sup>(2)</sup> | ETS(3)          |
|------------------------|-----------------|---------------------------------|-----------------|
| Echinochloa crus-galli | Low (0.4%)      | $62.3 \pm 11.7$                 | $1.52 \pm 0.19$ |
|                        | High (26%)      | $15.9 \pm 1.8$                  | $1.55 \pm 0.18$ |
| Setaria pumila         | Low (0.4%)      | $51.0 \pm 6.8$                  | $0.34 \pm 0.05$ |
|                        | High (26%)      | $28.0 \pm 5.6$                  | $0.32 \pm 0.06$ |
| Zea mays               | Low (0.1–0.4%)  | $239.0 \pm 21.0$                | $1.12 \pm 0.13$ |
|                        | High (over 10%) | $114.2 \pm 7.9$                 | $0.95 \pm 0.13$ |

<sup>&</sup>lt;sup>(1)</sup>Measured as soil CO<sub>2</sub> concentration (25 cm depth) by a gas analyzer GA 2000 (Ansyco, FRG). <sup>(2)</sup>By ANOVA, n = 10. <sup>(3)</sup>Given as  $\mu g O_2 g^{-1}$  fresh wt  $h^{-1}$ , by ANOVA, n = 12.

Results on *Echinochloa* and *Setaria*, could indicate a general low sensitivity of root respiration to a high  $CO_2$  concentration, which is suggested by LAMBERS et al. (1996, 2002). Yet, root respiratory potential in root tips of *Zea mays* measured in our study was significantly lower in the roots exposed to high soil  $CO_2$  than in those growing in the low  $CO_2$  environment (Tab. 2). Different results obtained for native species (*Echinochloa*, *Setaria*) and sown maize suggest that plants growing as a part of natural vegetation could be adapted to extreme conditions. This however, has to be confirmed in the future work.

### **Conclusions**

At natural CO<sub>2</sub> springs, the growth of *Echinochloa* and *Setaria* can be decreased by high CO<sub>2</sub> concentrations and physiological processes in shoots can be severely affected. Despite this, no significant impact of CO<sub>2</sub> exposure on root respiratory potential of the root-tip segments was found for the same species. More detailed physiological studies on root respiration are needed, regarding to the different parts of the root system, different ontogenetic development and measurements on different plant species. Further research is also needed in connection to different environmental factors affecting root respiration.

# Acknowledgements

Research was granted and performed in the frame of COST 627 program, granted by Forschungs-Pool 2001 (09112000) of the University of Essen and by grants J4-2186 and Z4-3196 from Ministry of Science, Education and Sport of Slovenia.

#### References

AMTHOR J. S. 1991: Respiration in a future, higher-CO<sub>2</sub> world. Plant, Cell and Environment 14: 13–20. Badiani M., Raschi A., Paolacci, A. R. & Miglietta F. 1999: Plants responses to elevated CO<sub>2</sub>; a perspective from natural CO<sub>2</sub> springs. In: AGRAWAL S. B. & AGRAWAL M. (ed.): Environmental Pollution And Plant Response, Lewis Pub., Boca Raton, pp. 45–81.

Buchanan B. B., Gruissem W. & Jones, R. L. (ed.) 2000: Biochemistry & molecular biology of plants. American Society of Plant Physiologists, Rockville, pp. 1177–1189.

- Burton A. J., Zogg G. P., Pregitzer K. S. & Zak D. R 1997: Effect of measurement CO<sub>2</sub> concentration on sugar maple root respiration. Tree Physiol. 17: 421–427.
- DRAKE B.G., GONZÀLEZ-MELER M.A. & LONG S.P. 1997: More efficient plants: a consequence of rising atmospheric CO<sub>2</sub>: Annu. Rev. Plant Physiol. Plant Mol. Biol. 48: 609–639.
- Ghannoum O., Von Caemmerer S., Ziska L. H. & Conroy J. P. 2000: The growth response of C4 plants to rising atmospheric CO<sub>2</sub> partial pressure: a reassessment. Plant, Cell and Environ 23: 931–942.
- KALIGARIČ M. 2001: Vegetation patterns and responses to elevated CO<sub>2</sub> from natural CO<sub>2</sub> springs at Strmec (Radenci, Slovenia). Acta Biologica Slovenica 44, 1-2: 31–38.
- Kenner A. A. & Ahmed S. I. 1975: Measurements of electron transport activities in marine phytoplankton. Mar. Biol. 33: 117-120.
- LAMBERS H., ATKIN O. K. & MILLENAAR F. F. 2002: Respiratory patterns in roots in relation to their functioning. In: Waisel Y., ESHEL A. & KAFKAFI U. (ed.): Plant Roots The Hidden Half, 3rd ed. Marcel Dekker, Inc., New York, pp. 521–552.
- Lambers H., Stulen I. & Van Der Werf, A. 1996: Carbon use in root respiration as affected by elevated atmospheric CO<sub>2</sub>. Plant and Soil, 187: 251–263.
- RASCHIA., MIGLIETTA F., TOGNETTI R. & VAN GARDINGEN P. R. (Ed.) 1997: Plant Responses to Elevated CO<sub>2</sub>. Cambridge University Press, Cambridge UK.
- TJOELKER M.G., OLEKSYN J., LEE, T.D. & REICH, P. B. 2001: Direct inhibition of leaf dark respiration by elevated CO<sub>2</sub> is minor in 12 grassland species. New Phytol. 150: 419–424.
- Turk B., Pfanz H., Vodnih D., Bernik R., Wittmann C., Sinkovič T. & Batič F. 2002: The effects of elevated CO<sub>2</sub> on bog rush (*Juncus effusus* L.) growing near natural CO<sub>2</sub> springs I. Effects on shoot anatomy. Phyton (Horn Austria) 42: 13–23.
- Vodnik D., Pfanz H., Maček I., Kastelec D., Lojen S. & Batič F. 2002: Photosynthetic performance of cockspur (*Echinochloa crus-galli* (L.) Beauv.) at sites of naturally elevated CO<sub>2</sub>. Photosynthetica 40(4): 575–579.
- Vodnik D., Pfanz H., Wittmann C., Maček I., Kastelec D., Turk B. & Batič F. 2002: Photosynthetic acclimation in plants growing near a carbon dioxide spring. Phyton (Horn Austria), 42: 239–244.
- VODNIK D., ŠIRCELJ H., KASTELEC D., MAČEK I., PFANZ H. & BATIČ F.: The effects of natural CO<sub>2</sub> enrichment on the growth of maize. Journal of Crop Production, in press.
- Yoder C. K., Vivib P., Defalco L. A., Seemann J. R. & Nowak R. S. 2000: Root growth and function of three Mojave Desert grasses in response to elevated atmospheric CO<sub>2</sub> concentration. New Phytol. 145: 245–256.
- Yoshioka T., Satoh S. & Yamasue Y. 1998: Effect of increased concentration of soil CO<sub>2</sub> on intermittent flushes of seed germination in *Echinochloa crus-galli* var. *crus-galli*. Plant, Cell and Environment 21: 1301–1306.
- ZISKA L. H. & BUNCE J. A. 1997: Influence of increasing carbon dioxide concentration on the photosynthetic and growth stimulation of selected C4 crops and weeds. Photosynth. Res. 54: 199–208.