Modulation of radiotherapy- and chemotherapy-induced normal tissue response as prophylaxis of their side effects

Pavlína Plevová

Department of Radiotherapy, University Hospital, Ostrava, Czech Republic

Background. Ionising radiation and cytostatic agents used in cancer therapy induce an immune response in normal tissues mediated by cytokines and adhesion molecules. Strategies modulating this response may downregulate cancer therapy side effects. The data published on the given topic have been reviewed. Conclusions. The strategies influencing the tissue immune response with the aim to reduce the side effects of chemotherapy and radiotherapy are conflicting. Some of them inhibit this response supposing that an exaggerated reaction may have a damaging effect (e.g. corticosteroids, nonsteroidal anti-inflammatory drugs (NSAID), lisofylline, anti-cytokine antibodies, anti-sense oligonucleotides, sialyl Lewis X analogues), others promote this reaction by inducing endogenous production of cytokines (AS101) or use recombinant forms of appropriate cytokines involved in this response in order to intensify the physiologic tissue response. In clinical practice, corticosteroids and NSAID are widely used to modulate this response, while other agents are still experimental.

Key words: radiotherapy – adverse effects; antineoplastic agents; antineoplastic agents – adverse effects; adjuvant, immunologic

Introduction

Ionising radiation and cytostatic agents used in cancer therapy exert damaging effects on normal tissues and induce there a complex response at the cellular and molecular levels. Cytokines and adhesion molecules are relea-

Received 25 February 2002 Accepted 11 March 2002

Correspondence to: Pavlína Plevová, M.D., Fr. Lyska 8, Ostrava-Belsky les, 700 30 Czech Republic, Phone: +420 69 6717841 or +420 69 6984370; Fax: +420 69 6919010; E-mail: pavlina.plevova@volny.cz

sed during this response and mediate intercellular interactions among the effectors of immune and other systems.^{1,2} Medical strategies that modulate this response in order to reduce chemotherapy- and radiotherapy-induced side effects are contradictory. Some of them inhibit this reaction, and their use is based on the hypothesis that exaggerated or persisting inflammatory response enhances the tissue damage; others stimulate this response in order to enhance physiological protective processes.

Acknowledgements: The author is indebted to Pavel Vodvářka for his helpful comments and Lenka Zivčáková for technical assistance.

a) Inhibition of the tissue response

Glucocorticoids exert strong anti-inflammatory effects including inhibition of pro-inflammatory cytokine production. 1-5 The molecular mechanism of their effects is not completely understood, but they inhibit the activity of some transcription factors.3 In clinical practice, corticosteroids are used to prevent or treat chemotherapy-induced nausea and vomitus⁶ and to prevent radiation- and chemotherapy-induced pneumonitis and fibrosis.^{7,8} Although corticosteroids suppressed radiation pneumonitis in an experimental model they were not able to reduce pulmonary fibrosis development. In another study, short-term use of dexamethasone suppressed temporarily radiation-induced pro-inflammatory cytokine gene expression in the mouse lung, but a rebound was observed after the drug withdrawal and the drug did little to change the essence and course of the pneumonitic process. 10 Dexamethasone is widely used in the prophylaxis of radiation-induced brain oedema and inflammation; this effect was demonstrated on an experimental model.^{7,11} Dexamethasone significantly reduces the incidence of the somnolence syndrome after prophylactic cranial irradiation in children with leukemia. 12 Betamethasone was beneficial in radiation-induced oral mucositis in a few patients.¹³ Dexamethasone delays the development of experimental radiation nephropathy; it does not stop the progression of injury. 14,15 Captopril, an angiotensin convertase enzyme inhibitor, enhanced the beneficial effect of dexamethasone in radiation nephropathy. 15 Corticosteroids suppress cytokine secretion in irradiated animal skin. 16 They reduce hematotoxic effects of 5-fluorouracil and methotrexate, but not of other cytostatic agents in an experimental model.¹⁷

Nonsteroidal anti-inflammatory drugs (NSAID) inhibit the prostaglandin synthesis through cyclooxygenase blockade, 18 activation of the transcription factor of nuclear factor κB (NF- κB) 19 and adhesion of neutrophils as a

result of a decreased expression of L-selectine. ¹⁸ In clinical practice, they are used in the treatment of fever, pain, and fatigue associated with chemotherapy and radiotherapy. Mesalazine has been studied in the prevention of oral mucositis; ²⁰ however, the result of this non-randomised study lacks clinical relevance. Indomethacine did not influence the survival of lethally irradiated mice. ²¹

Lisofylline is a xanthine derivative able to inhibit the release of various cytokines, such as TNF- α , TGF- β , MIP- 1α , IFN- γ , Il- 1β , IL-6, IL- $10.^{22,23}$ Its mechanism of action is thought to involve inhibition of acyl-substituted unsaturated phosphatidic acid, a second messenger lipid implicated in pro-inflammatory cytokine cellular activation. ^{24,25} It also decreases white cell adhesiveness. ²⁶ Lisofylline inhibited 5-fluorouracil-induced release of TGF- β and maybe also other hematopoiesis inhibiting cytokines and thus enhanced trilineage hematopoietic recovery after 5-fluorouracil treatment in mice. ²⁷

Pentoxifylline is a xanthine derivative with profound immunomodulatory properties in vitro, including inhibition of TNF- α , Il-1 β and IL-10 release. ^{23,28} Elevated levels of TNF- α have been shown to correlate with both the development and severity of transplantation-related complications. ²⁹ Although pentoxifylline reduced these complications in a study, ³⁰ these results were not reproduced in others including the one focused on 5-fluorouracilinduced oral mucositis. ³¹⁻³³

The results of a study with a TNF- α neutralising monoclonal antibody in transplant patients lack clinical significance.³⁴

Intravenous immunoglobulin, especially in high doses, has profound immunomodulatory effects, including the inhibition of anti-inflammatory cytokine release; 35,36 in addition, high TGF- β concentrations have been detected in intravenous immunoglobulin preparations. 37 Intravenous or intramuscular immunoglobulin has been studied sporadically in the prophylaxis or therapy of irradiation or

chemotherapy-induced oral mucositis and radiation pneumonitis. It is not possible to make a definite conclusion of its effects from these results.³⁸⁻⁴¹

As TGF- β plays an important role in the pathogenesis of fibrosis development, its inhibition might reduce the risk of this complication. Neutralising antibodies to both TGF-β₁ and TGF-β₂ significantly reduced the bleomycin-induced increase in the accumulation of lung collagen in an experimental model;42 However, fibrosis was ameliorated only partially.43 TGF-β antisense oligonucleotides, short synthetic deoxyribonucleotide oligomers complementary to DNA, prevent protein production.44 They have been investigated in the prevention of experimental peritoneal fibrous adhesions. 45 There are no reports on their use in association with chemotherapy or radiotherapy.

IL-4 has been shown to be able to downregulate radiation-induced production of mediators of inflammation, including IL1 β in the lung, suggesting its anti-inflammatory potential in regulating the radiation-induced response. ⁴⁶

Interferon γ , taurine, and niacin reduced bleomycin-induced pulmonary fibrosis in an animal model via TGF- β inhibition and subsequent procollagen expression downregulation. ⁴⁷⁻⁴⁹

The endothelial selectins (E-selectin and P-selectin) bind to sialylated tetrasaccharide sialyl Lewis X and A counter receptors on neutrophils, monocytes and lymphocytes, mediating their emigration into the tissue. ^{50,51} The analogues of sialyl Lewis X such as glycyrrhizin and carminic acid bind to E-selectin on irradiated endothelial cells and thereby inhibit adhesion of leukocytes and inflammatory response in vitro. ⁵²

b) Stimulation of the tissue response

It has been known for more than forty years that immunomodulators stimulating the cells of the reticulo-endothelial system can protect against deleterious effects of radiation.⁵³

AS 101 (ammonium trichloro (dioxyethylene-0,0') tellurate) stimulates some subpopulations of white cells and increases the release of various cytokines, including IL-1, IL-2, IL-6, TNF-α, GM-CSF, stem cell factor (SCF), and IFN-γ.⁵⁴⁻⁵⁷ AS 101 reduces hematotoxic effects of cyclophosphamide, 5-fluorouracil, doxorubicine, lomustine, carboplatin and etoposide, ⁵⁶⁻⁵⁹ and alopecia after carboplatin and etoposide. ⁵⁷ It also has been shown to exert radioprotective effects. ⁶⁰

The physiological role of cytokines in the immune response and tissue regeneration has led to experiments studying the effectiveness of recombinant forms of cytokines in the protection of normal tissues from damaging effects of chemotherapy and radiotherapy. The results of these experimental studies were successful, depending on the schedule and the dose of the cytokine used. 61,62

Recombinant IL-1 α , TNF- α , INF- γ administered before treatment reduced hematotoxic effects of both irradiation and chemotherapy with various agents. ^{21,62-70} G-CSF, GM-CSF, SCF act as radioprotectors both in vitro and in vivo; ⁷¹⁻⁷⁵ on the contrary, their concomitant administration with chemotherapy increases the sensitivity of hematopoietic cells to its cytotoxic effects. ⁷⁶ MIP-1 α exerts chemoprotective effects on bone marrow cells. ⁷⁷ IL-1 α also reduced small gut and lung toxicity of radio- or chemotherapy. ⁷⁸⁻⁸⁰

The combination of IL-1 and TNF- α had synergistic effects.⁶⁶ Both cytokines are relatively toxic due to their physiological roles in inflammation, especially after systemic application.⁸¹ G-CSF and GM-CSF are well tolerated and potentiate radioprotective effects of IL-1.⁶⁶

Local application of TGF-β3 on oral mucosa significantly reduced the 5-fluorouracil-induced oral mucositis in hamsters.^{82,83} IL-1, EGF, FGF have been shown to protect mice against ARA-C-induced alopecia.^{84,85}

The mechanism of the protective effects of cytokines might be explained by the following hypothesis: 1) Exogenous cytokines activate the physiological pathways of immune response through their receptors, thus activating and amplifying the defence of the organism. The induction of enzymes with antioxidant effects^{86,87} could be a part of this response. 2) Some cytokines, such as IL-1 or SCF, may directly or indirectly, through release of other cytokines, stimulate hematopoietic progenitor cells.61,62,74 3) Cytokines might inhibit the cell proliferation, thus reducing the sensitivity to proliferation-inhibiting agents or inducing the cell-cycling so that the cells enter into the relatively radio- or chemoresistent phases of the cell cycle, the S and G1 ones. 61,77 4) Certain cytokines, such as IL-6, IFN-γ, GM-CSF, inhibit cell apoptosis including its cytotoxic agents- and irradiation-induced activation.⁸⁸⁻⁹¹

Conclusions

The modulation of the tissue response to the damaging effects of radiotherapy and chemotherapy may reduce toxic effects of these treatment modalities. Only corticosteroids and NSAID are used in clinical practice to reduce acute toxicity of cancer therapy. The agents that could affect late sequels are studied experimentally; AS101 is being tested at the clinical level. The response-modifying use of recombinant cytokines to reduce toxicity of radiotherapy or chemotherapy did not progress into clinical usage. The local use of TGF- β in association with chemotherapy-induced oral mucositis is promising.

The suppression of the inflammatory response must be used with caution in the clinical practice, however. Although corticosteroids are beneficial in the modulation of acute side effects, this effect results from inhibition of the protective response that is of pivotal importance in the maintenance of organism integrity and whose suppression might have

detrimental end-effects as has been demonstrated by reduced survival of mice that were administered dexamethasone after irradiation. 92,93

References

- Thalmeier K, Meissner P, Reisbach G, Hültner L, Mortensen BT, Brechtel A, et al. Constitutive and modulated cytokine expression in two permanent human bone marrow stromal cell lines. *Exp Hema*tol 1996; 24: 1-10.
- Koj A. Initiation of acute phase response and synthesis of cytokines. *Biochim Biophys Acta* 1996; 1317: 84-94.
- Barnes PJ, Karin M. Nuclear factor-κB a pivotal transcription factor in chronic inflammatory diseases. N Engl J Med 1997; 336: 1066-71.
- Kerner B, Teichmann B, Welte K. Dexamethasone inhibits tumor necrosis factor-induced granulocyte colony-stimulating factor production in human endothelial cells. Exp Hematol 1992; 20: 334-8.
- Tobler A, Meier R, Seitz M, Dewald B, Baggiolini M, Fey MF. Glucocorticoids downregulate gene expression of GM-CSF, NAP-1/IL-8, and IL-6 but not of M-CSF in human fibroblasts. *Blood* 1992; 79: 45-51.
- The Antiemetic Subcommittee of the Multinational Association of Supportive Care in Cancer (MASCC). Prevention of chemotherapy- and radiotherapy-induced emesis: results of Perugia Consensus Conference. *Ann Oncol* 1998; 9: 811-9.
- Rubin P, Constine LS, Williams JP. Late effects of cancer treatment: radiation and drug toxicity. In: Perez CA, Brady LW, editors. Principles and practice of radiation oncology. 3rd ed. Philadelphia: Lippincott-Raven Publishers; 1998. p. 155-212.
- 8. Khan A, McNally D, Tutschka PJ, Bilgrami S. Paclitaxel-induced acute bilateral pneumonitis. *Ann Pharmacother* 1997; **31:** 1471-4.
- Ward HE, Kemsley L, Davies L, Holecek M, Berend N. The effect of steroids on radiation-induced lung disease in the rat. *Radiat Res* 1993; 136: 22-8.
- Hong JH, Chiang CS, Tsao CY, Lin PY, Wu CJ, McBride WH. Can short-term administration of dexamethasone abrogate radiation-induced acute cytokine gene response in lung and modify subsequent molecular responses? *Int J Radiat Oncol Biol Phys* 2001; 51: 296-303.

- Tada E, Matsumoto K, Kinoshita K, Furuta T, Ohmoto T. The protective effect of dexamethasone against radiation damage induced by interstitial irradiation in normal monkey brain. *Neurosur*gery 1997; 41: 209-19.
- Uzal D, Ozyar E, Hayran M, Zorlu F, Atahan L, Yetkin S. Reduced incidence of the somnolence syndrome after prohyplactic cranial irradiation in children with acute lymphoblastic leukemia. *Radiother Oncol* 1998; 48: 29-32.
- 13. Abdelaal AS, Barker DS, Fergusson MM. Treatment for irradiation-induced mucositis [letter]. *Lancet* 1989; **1**: 97.
- 14. Robbins ME, Bonsib SM. Radiation nephropathy: a review. *Scanning Microsc* 1995; **9:** 535-60.
- Geraci JP, Sun MC, Mariano MS. Amelioration of radiation nephropathy in rats by postirradiation treatment with dexamethasone and/or captopril. *Radiat Res* 1995; 143: 58-68.
- Beetz A, Messer G, Oppel T, van Beuningen D, Peter RU, Kind P. Induction of interleukin 6 by ionizing radiation in a human epithelial cell line: control by corticosteroids. *Int J Radiat Biol* 1997; 72: 33-43.
- Kriegler AB, Bernardo D, Verschoor SM. Protection of murine bone marrow by dexamethasone during cytotoxic chemotherapy. *Blood* 1994; 83: 65-71.
- Díaz-González F, Sánchez-Madrid F. Inhibition of leukocyte adhesion: an alternative mechanism of action for anti-inflammatory drugs. *Immunol Today* 1998; 19: 169-72.
- Pierce JW, Read MA, Ding H, Luscinskas FW, Collins T. Salicylates inhibit I kappa B-α phosphorylation, endothelial-leukocyte adhesion molecule expression and neutrophil transmigration. *J Immunol* 1996; 156: 3961-9.
- Rymes N, Glick L, Holmes JA. Topical mesalazine in the treatment of chemotherapy and radiotherapy-induced oral mucositis [letter]. *Bone Marrow Transplant* 1996; 18: 484.
- 21. Neta R. Role of cytokines in radioprotection. *Pharmacol Ther* 1988; **39:** 261-6.
- Rice GC, Rosen J, Weeks R, Michnick J, Bursten S, Bianco JA, et al. CT-1501R selectively inhibits induced inflammatory monokines in human whole blood ex vivo. Shock 1994; 1: 254-66.
- van Furth AM, Verhard-Seijmonsbergen EM, van Furth R, Langermans JA. Effect of lisofylline and pentoxyphylline on the bacterial-stimulated production of TNF-α, II-1β, IL-10 by human leucocytes. *Immunology* 1997; 91: 193-6.

- 24. Bursten S, Weeks R, West J, Le T, Wilson T, Porubek D, et al. A potential role for phosphatidic acid in mediating the inflammatory responses of TNF and IL-1. *Circ Shoc* 1994; 44: 14-29.
- Schwaighofer H, Kernan NA, O'Reilly RJ, Brankova J, Nachbaur D, Herold M, et al. Serum levels of cytokines and secondary messages after T-cell-depleted and non-T-cell-depleted bone marrow transplantation: influence of conditioning and hematopoietic reconstitution. *Transplantation* 1996; 62: 947-53.
- Waxman K, Daughters K, Aswani S, Rice G. Lisofylline decreases white cell adhesiveness and improves survival after experimental hemorrhagic shock. Crit Care Med 1996; 24: 1724-8.
- 27. Clarke E, Rice GC, Weeks RS, Jenkins N, Nelson R, Bianco JA, et al. Lisofylline inhibits transforming growth factor β release and enhances trilineage hematopoietic recovery after 5-fluorouracil treatment in mice. Cancer Res 1996; 56: 105-12.
- Tilg H, Eibl B, Pichl M, Gachter A, Herold M, Brankova J, et al. Immune response modulation by pentoxifylline in vitro. *Transplantation* 1993; 56: 196-201.
- Holler E, Kolb HJ, Möller A, Kempeni J, Liesenfeld S, Pechumer H, et al. Increased serum levels of tumor necrosis factor α precede major complications of bone marrow transplantation. *Blood* 1990; 75: 1011-6.
- Bianco JA, Appelbaum FR, Nemunaitis J, Almgren J, Andrews F, Kettner P, et al. Phase I-II trial of pentoxifylline for the prevention of transplant-related toxicities following bone marrow transplantation. *Blood* 1991; 78: 1205-11.
- Clift RA, Bianco JA, Appelbaum FR, Buckner CD, Singer JW, Bakke L, et al. A randomized controlled trial of pentoxifylline for the prevention of regimen-related toxicities in patients undergoing allogeneic marrow transplantation. *Blood* 1993; 82: 2025-30.
- 32. Attal M, Huguet F, Rubie H, Charlet JP, Schlaifer D, Huynh A, et al. Prevention of regimen-related toxicities after bone marrow transplantation by pentoxifylline: A prospective, randomized trial. Blood 1993; 82: 732-6.
- 33. van der Jagt RHC, Pari G, McDiarmid SA, Boisvert DM, Huebsch LB. Effect of pentoxifyline on regimen related toxicity in patients undergoing allogeneic or autologous bone marrow transplantation. Bone Marrow Transplant 1994; 13: 203-7.
- 34. Holler E, Kolb HJ, Mittermüller J, Kaul M, Ledderose G, Duell T, et al. Modulation of acute graft-versus-host disease after allogeneic bone marrow

- transplantation by tumor necrosis factor α (TNF α) release in the course of pretransplant conditioning: Role of conditioning regimens and prophylactic application of a monoclonal antibody neutralizing human TNF α (MAK 195F). *Blood* 1995; **86:** 890-9.
- 35. Nydegger U. [Old and new views on intravenous immunoglobulin therapy.] *Schweiz Med Wochenschr* 1994; **124:** 5-25 (German).
- Wolf HM, Eibl MM. Immunomodulatory effect of immunoglobulins. Clin Exp Rheumatol 1996; 14 (Suppl 15): S17-25.
- 37. Kekow J, Reinhold D, Pap T, Ansorge S. Intravenous immunoglobulins and transforming growth factor β. *Lancet* 1998; **351**: 184-5.
- 38. Proske H, Pfab R. [Immunoglobulin preparations as antiinflammatory drugs in radiotherapy]. *Med Welt* 1992; **43**: 1025-26 (German).
- 39. Schedler MGJ, Bost P, Federspil P, Pautler M, Schatzle W. Treatment of radiogenic mucositis in patients with head and neck tumors with polyvalent intramuscular immunoglobulin. *Tumor Diagn Ther* 1994; 15:184-91 (German).
- Plevová P, Blažek B. Intravenous immunoglobulin as prophylaxis of chemotherapy-induced oral mucositis. J Natl Cancer Inst 1997; 89: 326-7.
- 41. Seibel RM, Wendt BK. Immunoglobulin as prophylaxis of ionizing-radiation induced pneumonitis following high-volume irradiation for lung cancer. *Onkologie* 1986; 9: 43-7 (German).
- Giri SN, Hyde DM, Hollinger MA. Effect of antibody to transforming growth factor β on bleomycin induced accumulation of lung collagen in mice. *Thorax* 1993; 48: 959-66.
- Laurent GJ, Coker RK, McAnulty RJ. TGF-beta antibodies: a novel treatment for pulmonary fibrosis? *Thorax* 1993; 48: 953-4.
- 44. Khanna A, Li B, Li P, Suthanthiran M. Regulation of transforming growth factor-beta 1 (TGF-β1) expression with a novel TGF-β1 complementary DNA. *Biochem Biophys Res Commun* 1994; **204**: 1061-6.
- 45. Chegini N. The role of growth factors in peritoneal healing: transforming growth factor β (TGF-β). *Eur J Surg* 1997; **577(Suppl):** 17-23.
- 46. Van der Meeren A, Monti P, Lebaron-Jacobs L, Marquette C, Gourmelon P. Characterization of the acute inflammatory response after irradiation in mice and its regulation by interleukin 4 (II4). Radiat Res 2001; 155: 858-65.

- 47. Gurujeyalakshmi G, Giri SN. Molecular mechanisms of antifibrotic effect of interferon gamma in bleomycin-mouse model of lung fibrosis: downregulation of TGF-β and procollagen I and III gene expresssion. *Exp Lung Res* 1995; **21**: 791-808.
- 48. Gurujeyalakshmi G, Hollinger MA, Giri SN. Regulation of transforming growth factor-β1 mRNA epression by taurine and niacin in the bleomycin hamster model of lung fibrosis. *Am J Respir Cell Mol Biol* 1998; **18:** 334-42.
- Gurujeyalakshmi G, Iyer SN, Hollinger MA, Giri SN. Procollagen gene expression is down-regulated by taurine and niacin at the transcriptional level in the bleomycin hamster model of lung fibrosis. J Pharmacol Exp Ther 1996; 277: 1152-7.
- 50. Rosen SD. Cell surface lectins in the immune system. *Semin Immunol* 1993; **65:** 237-47.
- 51. Lasky LA. Selectins: interpreters of cell-specific carbohydrate information during inflammation. *Science* 1992; **258**: 964-9.
- 52. Hallahan DE, Kuchibhotla J, Wyble C. Sialyl Lewis X mimetics attenuate E-selectin-mediated adhesion of leukocytes to irradiated human endothelial cells. *Radiat Res* 1997; **147**: 41-7.
- Mefferd RB, Henkel DT, Loeffer JB. Effect of piromen on survival of irradiated mice. Proc Soc Exp Biol Med 1953; 83: 54-6.
- 54. Sredni B, Caspi RR, Lustig S, Klein A, Kalechman Y, Danzinger Y, et al. The biological activity and immunotherapeutic properties of AS-101, a synthetic organotellurium compound. *Nat Immun Cell Growth Regul* 1988; 7: 163-8.
- 55. Shani A, Gurwith M, Tichler T, Catane R, Rozenszajan LA, Gezin A, et al. The immunologic effects of AS 101 in the treatment of cancer patients. *Nat Immun Cell Growth Regul* 1990; 9: 182-90.
- Kalechman Y, Zuloff A, Albeck M, Strassmann G, Sredni B. Role of endogenous cytokines secretion in radioprotection conferred by the immunomodulator ammonium trichloro(dioxyethylene-0-0')tellurate. *Blood* 1995; 85: 1555-61.
- 57. Sredni B, Albeck M, Tichler T, Shani A, Shapira J, Bruderman I, et al. Bone marrow-sparing and prevention of alopecia by AS 101 in non-small-cell lung cancer patients treated with carboplatin and etoposide. *J Clin Oncol* 1995; 13: 2342-53.
- 58. Kalechman Y, Sotnik-Barkai I, Albeck M, Sredni B. Protection of bone marrow stromal cells from the toxic effects of cyclophosphamide in vivo and of ASTA-Z 7557 and etoposide in vitro by ammonium trichloro(dioxyethylene-0-0') tellurate (AS 101). Cancer Res 1993; 53: 1838-44.

- Kalechman Y, Shani A, Sotnik Barkai I, Albeck M, Sredni B. The protective role of ammonium trichloro(dioxyethylene-0,0)tellurate in combination with several cytotoxic drugs acting by different mechanisms of action. *Cancer Res* 1993; 53: 5962-9.
- Kalechman Y, Albeck M, Oron M, Sobelman D, Gurwith M, Sehgal SN, et al. Radioprotective effects of the immunomodulator AS 101. *J Immunol* 1990; 145: 1512-7.
- Dalmau SR, Freitas CS, Tabak DG. Interleukin -1 and tumor necrosis factor-alpha as radio- and chemoprotectors of bone marrow. *Bone Marrow Trans*plant 1993; 12: 551-63.
- 62. Wu SG, Tuboi A, Miyamoto T. Radioprotection of C3H mice by recombinant human interleukin-1 alpha. *Int J Radiat Biol* 1989; **56**: 485-92.
- 63. Neta R, Keller JR, Ali N, Blanchette F, Dubois CM. Contrasting mechanisms of the myeloprotective effects of interleukin-1 against ionizing radiation and cytotoxic 5-fluorouracil. *Radiat Res* 1996; 145: 624-31.
- 64. Neta R, Sztein MB, Oppenheim JJ, Gillis S, Douches SD. The in vivo effects of interleukin 1: I. Bone marrow cells are induced to cycle after administration of interleukin 1. *J Immunol* 1987; 139: 1861-6.
- Castelli MP, Black PL, Schneider M, Pennington R, Abe F, Talmadge JE. Protective, restorative and therapeutic properties of recombinant human IL-1 in rodent models. J Immunol 1988; 140: 3830-7.
- 66. Neta R, Oppenheim JJ, Douches SD. Interdependence of the radioprotective effects of human recombinant interleukin 1, tumor necrosis factor, granulocyte colony-stimulating factor, and murine recombinant granulocyte-macrophage colony-stimulating factor. J Immunol 1988; 140: 108-11.
- 67. Futami H, Jansen R, MacPhee MJ, Keller JR, McCormick K, Longo DL, et al. Chemoprotective effects of recombinant human IL-1α in normal and tumor-bearing mice. Protection from acute toxicity, hematologic effects, development of late mortality and enhanced therapeutic efficacy. J Immunol 1990; 145: 4121-30.
- Slordal L, Warren DJ, Moore MAS. Effect of recombinant murine tumor necrosis factor on hemopoietic reconstitution in sublethally irradiated mice. J Immunol 1989; 142: 833-5.
- Slordal L, Warren DJ, Moore MAS. Protective effects of tumor necrosis factor on murine hematopoiesis during cycle-specific cytotoxic chemotherapy. *Cancer Res* 1990; 50: 4216-20.

- 70. Gardner RV. Interferon-gamma (IFN-γ) as a potential radio- and chemoprotectant. *Am J Hematol* 1998; **58**: 218-23.
- Uckun FM, Gillis S, Souza L, Song CW. Effects of recombinant growth factors on radiation survival of human bone marrow progenitor cells. *Int J Radiat Oncol Biol Phys* 1989; 16: 415-35.
- Uckun FM, Souza L, Waddick KG, Wick M, Song CW. In vivo radioprotective effects of recombinant human granulocyte colony-stimulating factor in lethally irradiated mice. *Blood* 1990; 75: 638-45.
- Waddick KG, Song CW, Souza L, Uckun FM. Comparative analysis of the in vivo radioprotective effects of recombinant granulocyte colony-stimulating factor (G-CSF), recombinant granulocyte-macrophage CSF, and their combination. *Blood* 1991; 77: 2364-71.
- Zsebo KM, Smith KA, Hartley CA, Greenblatt M, Cooke K, Rich W, et al. Radioprotection of mice by recombinant rat stem cell factor. *Proc Natl Acad Sci* USA 1992; 89: 9464-8.
- 75. Liebmann J, DeLuca AM, Epstein A, Steinberg SM, Morstyn G, Mitchtell JB. Protection from lethal irradiation by the combination of stem cell factor and tempol. *Radiat Res* 1994; **137**: 400-4.
- Meropol NJ, Miller LL, Korn EL, Braitman LE, MacDermott ML, Schuchter LM. Severe myelosuppression resulting from concurrent administration of granulocyte colony-stimulating factor and cytotoxic chemotherapy. J Natl Cancer Inst 1992; 84: 1201-3.
- Dalmau SR, Freitas CS, Savino W. Radio- and chemoprotection of bone marrow cells by opposite cell cycle-acting cytokines. *Leuk Res* 1997; 21: 93-9.
- 78. Neta R, Douches S, Oppenheim JJ. Interleukin 1 is a radioprotector. *J Immunol 1986*; **136**: 2483-5.
- Dorie MJ. Protection by interleukin 1 against lung toxicity caused by cyclophosphamide and irradiation. *Radiat Res* 1991; 128: 316-9.
- Damia G, Komschlies KL, Futami H, Back T, Gruys ME, Longo DL, et al. Prevention of acute chemotherapy-induced death in mice by recombinant human interleukin 1: protection from hematological and nonhematological toxicities. *Cancer Res* 1992; 52: 4082-9.
- Maisin JR. Bacq and Alexander Award Lecture. Chemical radioprotection: past, present and future prospects. *Int J Radiat Biol* 1998; 73: 443-50.
- 82. Sonis ST, Lindquist L, van Vugt V, Stewart AA, Stam K, Qu GY, et al: Prevention of chemothe-

- rapy-induced ulcerative mucositis by transforming growth factor β 3. *Cancer Res* 1994; **54**: 1135-8.
- 83. Sonis ST, van Vugt AG, Brien JP, Muska AD, Bruskin AM, Rose A, et al. Transforming growth factor-β3 mediated modulation of cell cycling and attenuation of 5-fluorouracil induced oral mucositis. *Oral Oncol* 1997: 33: 47-54.
- 84. Jimenez JJ, Wong GH, Yunis AA. Interleukin 1 protects from cytosine arabinoside-induced alopecia in the rat model. *FASEB J* 1991; **5:** 2456-8.
- 85. Jimenez JJ, Yunis AA. Protection from 1-beta-Darabinofuranosylcytosine-induced alopecia by epidermal growth factor and fibroblast growth factor in the rat model. *Cancer Res* 1992; **52**: 413-5.
- 86. Masuda A, Longo DL, Kobayashi Y, Appella E, Oppenheim JJ, Matsushima K. Induction of mitochondrial manganese superoxide dismutase by interleukin 1. *FASEB J* 1988; **2:** 3087-91.
- 87. Wong GHW, Goeddel DV. Induction of manganous superoxide dismutase by tumor necrosis factor: possible protective mechanism. *Science* 1988; **242**: 941-4.
- Lotem J, Sachs L. Differential suppression by protease inhibitors and cytokines of apoptosis induced by wild-type p53 and cytotoxic agents. *Proc* Natl Acad Sci USA 1996; 93: 12507-12.

- Lotem J, Sachs L. Control of apoptosis in hematopoiesis and leukemia by cytokines, tumor suppressor and oncogenes. *Leukemia* 1996; 10: 925-31.
- Lotem J, Sachs L. Cytokine suppression of protease activation in wild-type p53-dependent and p53independent apoptosis. *Proc Natl Acad Sci USA* 1997: 94: 9349-53.
- Mor F, Cohen IR. IL-2 rescues antigen-specific T cells from radiation or dexamethasone-induced apoptosis. Correlation with induction of Bcl-2. *J Immunol* 1996; 156: 515-22.
- 92. Nam SY, Cho CK, Kim SG. Correlation of increased mortality with the suppression of radiation-inducible microsomal epoxide hydrolase and glutathione D-transferase gene expression by dexamethasone: effects on vitamin C and E-induced radioprotection. *Biochem Pharmacol* 1998; 56: 1295-34.
- 93. Rudat V, Kupper JH, Weber KJ. Trans-dominant inhibition of poly(ADP-ribosyl)ation leads to decreased recovery from ionizing radiation-induced cell killing. *Int J Radiat Biol* 1998; **73**: 325-30.